



Review

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Authors for correspondence:

Jonci N. Wolff

e-mail: jonciwolff@yahoo.com

Damian K. Dowling

e-mail: damian.dowling@monash.edu.au

Mitonuclear interactions: evolutionary consequences over multiple biological scales

Jonci N. Wolff^{1,2,6}, Emmanuel D. Ladoukakis³, José A. Enríquez^{4,5}
and Damian K. Dowling⁶

¹School of Biotechnology and Biomolecular Sciences, and ²Evolution and Ecology Research Centre, University of New South Wales, Sydney 2052, New South Wales, Australia

³Department of Biology, University of Crete, 70013 Heraklion, Crete, Greece

⁴Regenerative Cardiology Department, Centro Nacional de Investigaciones Cardiovasculares Carlos III, Madrid, Spain

⁵Departamento de Bioquímica, Universidad de Zaragoza, Zaragoza, Spain

⁶School of Biological Sciences, Monash University, Clayton 3800, Victoria, Australia

Fundamental biological processes hinge on coordinated interactions between genes spanning two obligate genomes—mitochondrial and nuclear. These interactions are key to complex life, and allelic variation that accumulates and persists at the loci embroiled in such intergenomic interactions should therefore be subjected to intense selection to maintain integrity of the mitochondrial electron transport system. Here, we compile evidence that suggests that mitochondrial–nuclear (mitonuclear) allelic interactions are evolutionarily significant modulators of the expression of key health-related and life-history phenotypes, across several biological scales—within species (intra- and interpopulational) and between species. We then introduce a new frontier for the study of mitonuclear interactions—those that occur within individuals, and are fuelled by the mtDNA heteroplasmy and the existence of nuclear-encoded mitochondrial gene duplicates and isoforms. Empirical evidence supports the idea of high-resolution tissue- and environment-specific modulation of intraindividual mitonuclear interactions. Predicting the penetrance, severity and expression patterns of mtDNA-induced mitochondrial diseases remains a conundrum. We contend that a deeper understanding of the dynamics and ramifications of mitonuclear interactions, across all biological levels, will provide key insights that tangibly advance our understanding, not only of core evolutionary processes, but also of the complex genetics underlying human mitochondrial disease.

1. Introduction

One of life's most important biological functions—ATP production via oxidative phosphorylation (OXPHOS)—critically depends on the concerted and synchronized interaction between genes encoded over two obligate genomes—nuclear and mitochondrial. Respiratory function and its regulation in response to cellular and environmental cues (e.g. nutrients, exercise, temperature, hypoxia, etc.) necessitates the recruitment of over 500 proteins into the mitochondria [1–4]. The very essence of complex eukaryotic life depends on the interactions that take place between these imported nuclear-encoded proteins, and those encoded by the mitochondrial DNA (mtDNA) [5]. At its core, therefore, the mitochondrial–nuclear (mitonuclear) genomic interaction is essential for the assembly and function of the mitochondrial electron transport system (mETS). The mETS is the site of OXPHOS, and in eukaryotes entails five multi-subunit enzyme complexes, four of which are composed of subunits encoded by both mitochondrial (13 subunits) and nuclear (approx. 73 subunits) genomes [6]. Protein subunits from the two genomes must be highly compatible—akin to a 'lock and key' principle—in order to maintain structural and biochemical properties to ensure uncompromised enzymatic function.

The genetic architecture (in terms of genes involved), and genomic location (in terms of in which genome each gene resides), of the genes that encode subunits of the mETS are remarkably conserved across metazoans [7]. What is, however, subjected to considerable heterogeneity is the level of nucleotide variation observed across these OXPHOS-coding genes. In particular, variation in the mtDNA sequence has long been known to exist over several biological scales—between congeneric species, between populations of the same species, within panmictic populations, and even within individuals. With the exception of the intraindividual level of variation [8], this sequence variation was historically neglected in terms of its functional and evolutionary relevance. Rather, the variation found within the sequence was typically considered to accumulate under a neutral equilibrium model, by-and-large silent to selection, and with little if any relevance as a mediator of population evolutionary processes.

The past two decades have seen a substantial shift away from this traditional view. This shift is founded on accumulating evidence, spanning vertebrate and invertebrate model systems, which indicates that the sequence polymorphism observed in mtDNA is ubiquitously tied to sizeable phenotypic effects [9–13]. Much of this evidence comes from studies that have associated putatively healthy mitochondrial genetic variants to phenotypic variation [14–22]. Other evidence comes from the medical sciences, where studies have reported associations between mutations in the human mitochondrial genome and deficiencies in mitochondrial function and the expression of disease [8,23]. In certain cases, disease phenotypes can be traced to specific mitochondrial polymorphisms (e.g. mitochondrial encephalomyopathy, lactic acidosis and stroke-like episodes [24] or Leber hereditary optic neuropathy (LHON) [25]). In many cases, however, while particular diseases are linked to mitochondrial function (e.g. diabetes [26], Alzheimer's [27], Parkinson's [28], cancer [29,30], ageing [31–33], etc.), direct links to specific disease-causing mtDNA polymorphisms remain elusive, a shortcoming further complicated by the varying nature, penetrance and severity of mitochondrial degenerative diseases [8,32]. We contend that a non-trivial amount of the currently unexplained variance in penetrance and expression of mitochondrial diseases might ultimately be traced to the unit of the mitonuclear interaction.

2. Mitonuclear interactions and coevolutionary processes

The mitonuclear interaction governs the integrity of OXPHOS and ensuing metabolic functions, and therefore should be under intense selection. All else being equal, deleterious mutations that arise in either genome should be swiftly eliminated to ensure uncompromised function, and selection should optimize combinations of alleles located on mitochondrial and nuclear genomes that work particularly well together, resulting in these combinations increasing in frequency within populations over evolutionary time [10]. Because the pool of genetic variation—created through de novo mutations in each genome, and the standing genetic variation—will differ across populations, and because each population is exposed to differing regimes of natural selection as a result of inhabiting distinct spatial and temporal

environments, coevolutionary trajectories of mitonuclear coadaptation are predicted to be population-specific.

Yet, the general dynamics of mitonuclear interactions manifested in nature, and modes of ensuing coevolution, remain enigmatic and largely uncharacterized [10,34]. This shortcoming is most likely fuelled by different tempi and modes at which the two genomes evolve, requiring swift and efficient selective processes to fix or purge allelic variation arising in either genome to uphold OXPHOS function. The simple verbal model discussed above, whereby coevolution involves adaptations selected over both genomes to promote optimized function, is certainly plausible. However, several asymmetries in the genetics of the mitochondrial and nuclear genomes would suggest that the adaptive component of mitonuclear coevolution might commonly be left in the hands of the nuclear genome [10,34]. In particular, the mitochondrial genome is thought to have a small effective population size (N_e) as a result of its maternal inheritance and haploidy, relative to its nuclear counterpart [35]. The implication is that the efficacy of selection in shaping the mtDNA sequence across generations should be dampened, and the effects of genetic drift amplified [10,12,36]. When coupled with absence of recombination [37, but see 38–40] and the observations that mutation rates within the metazoan mitochondrial genomes are typically high, then it can be expected that non-adaptive processes will play a pervasive role in driving mitochondrial genome evolution, and facilitate the perpetual accumulation of deleterious mutations within mtDNA sequences [36,41]. The accumulation of mtDNA mutations, which threaten OXPHOS integrity and associated phenotypes, should therefore place intense selection on the nuclear genome for counter-adaptations that restore compromised function. This model describes a coevolutionary process that is primarily compensatory—where the nuclear genome must repeatedly come to the evolutionary rescue of mtDNA-induced mitochondrial dysfunction [10,34], a notion congruent with the genetic footprint of compensatory adaptation observable in nuclear genes encoding subunits of the cytochrome *c* oxidase complex in primates [42].

While the relative contributions of adaptive to non-adaptive processes in shaping mitochondrial genome evolution are unclear, it is likely that each of the coevolutionary models (co-adaptive and compensatory) described above contribute to shaping the genomic landscape of the mitonuclear genomic interaction. And regardless of the specific drivers involved, we predict that the footprint of mitonuclear coevolution will be significant, and leave its mark over multiple taxonomic scales. Evolutionary forces acting within populations will drive population-specific evolutionary trajectories of mitonuclear genotypes, which over evolutionary time will promote reproductive isolation caused by mitonuclear allelic incompatibilities between disjunct populations, with these incompatibilities ultimately serving as engines of speciation [43].

In this paper, we outline evidence that suggests that the expression of key health parameters, and life-history phenotypes, are routinely shaped by mitonuclear interactions. Furthermore, these interactions are concurrently taking place, and are evolutionarily relevant, over several tiers of biological structure (intrapopulation to interpopulation, and interspecies). We argue that a deeper understanding of the ubiquity and magnitude of these interactions has potential to feed into our general understanding of core evolutionary concepts, and to also provide insights into the complex genetic basis of

mitochondrial diseases. We then conclude by introducing a fourth tier on which mitonuclear interactions could well exert significant effects, but remain completely unstudied—the ‘intra-individual’ tier. Individuals often carry heteroplasmic mtDNA sequences (a non-clonal population of mitochondrial genomes, typically consisting of wild-type and mutant mtDNA molecules) [44,45]. Furthermore, many nuclear-encoded mitochondrial genes are also present in duplicates, and these duplicates often exhibit tissue-specific expression [46–48]. Thus, allelic variation within particular mtDNA- and nuclear-encoded genes can co-segregate, and thus potentially interact, even within an individual, and this might have hitherto unrealized medical and evolutionary implications.

3. Intrapopulation interactions

Given the haploid nature of the mitochondrial genome, all de novo mutations appearing in the mtDNA will be continually exposed to the full force of natural selection. When this observation is coupled with the knowledge that the mtDNA encodes some of life’s most essential gene products, one could conclude that directional selection will generally efficiently purge (when pathogenic) or fix (when beneficial) any phenotype-changing (i.e. functional) mutations, contingent on the magnitude of their associated effects [10]. This would suggest, *prima facie*, that most polymorphisms segregating in the mitochondrial genome of metazoan species are likely to be selectively neutral, or near-neutral. However, as discussed above, the diminished efficacy of selection in shaping the mitochondrial genome that directly results from a low N_e , should confer an upward shift in the equilibrium frequencies at which deleterious mutations are maintained within the mitochondrial genome, under mutation–selection balance, relative to that which might be expected for mutations of equivalent effect in the nuclear genome.

Theoreticians have addressed the potential for stable joint polymorphisms in the mitochondrial and nuclear genomes to be maintained within populations via mitonuclear interactions (a form of balancing selection) [49–53]. These studies have shown a range of conditions under which protected mitonuclear or mitochondrial polymorphisms could be upheld, such as under frequency-dependent or sex-specific selection [49], or sex linkage of the interacting nuclear genes [50]. Based on empirical results derived from the interpopulation level, other authors have suggested that gene-by-environment interactions involving polymorphisms in the mtDNA, including complex mtDNA-by-nuclear-by-environment effects, could plausibly uphold mitochondrial genetic variance within populations [54,55]. Relevant empirical work at the intrapopulation scale is scarce [50,54,56,57], and all comes from work on *Drosophila melanogaster*. These studies generally did not have the required resolution to home their evidence of cytonuclear interactions to the level of the mtDNA sequence, but they nonetheless provide some proof of concept for the idea that mtDNA polymorphisms could be maintained within populations via mitonuclear interactions for fitness [50,54].

Much research remains to be done at the intrapopulation level to empirically validate the possibility and extent to which polymorphisms in mtDNA are maintained by mitonuclear interactions. To date, the few studies conducted at the intrapopulation scale would suggest that the links between

the mitochondrial genotype and phenotype are highly contingent on the particular nuclear background with which the mtDNA is coexpressed. These results give credence to the hypothesis that mitonuclear allelic variation can be maintained under adaptive balancing selection. Thus, selection may act to preserve levels of functional mitochondrial genetic variance in nature, and to maintain multiple mitonuclear allelic combinations, which regulate OXPHOS function. We believe that this might be important to long-term sustainability of our biodiversity in the face of directional change in environmental parameters such as temperature, and the creation of novel environmental, thus, selection pressures. The existence of functional allelic variation underlying OXPHOS function, which relies on uncompromised function of highly temperature-sensitive enzyme complexes, might provide the fuel on which populations can mount adaptive evolutionary responses to preserve their metabolic capacity during bouts of sustained changes to their environments.

4. Interpopulation interactions

In the absence of high levels of gene flow, conspecific populations are expected to coevolve along population-specific trajectories, with each population accumulating its own set of adaptations in interacting regions of each of the mtDNA and nuclear DNA, as well as deleterious mutations that arise in the mtDNA, and their nuclear restorer alleles. The result is that mitonuclear interactions—and OXPHOS function—will evolve towards population-specific optima, and the mitonuclear componentry of incipient populations may become increasingly incompatible as populations diverge, under a Dobzhansky–Muller model of hybrid incompatibilities (DMIs) [58–60] in which the mitonuclear unit takes centre stage [61]. Considering the fundamental role of OXPHOS function to components of organismal fitness, any disruption to the coevolved mitonuclear allelic combination is thus expected to invoke deleterious fitness consequences.

Recent evidence supports the notion that tight mitonuclear coevolution underpins organismal fitness. This has been most prominently illustrated by observations of intergenomic incompatibilities leading to mitochondrial dysfunction following experimental hybridization in the marine copepod, *Tigriopus californicus*. Such hybridization results in fitness breakdown in the F₂ generation, marked by decreased survivorship of larvae [62], slower development [63], reduced fecundity and viability [64], as well as decreased cyclooxygenase (COX) activity and ATP production [64–66]. The decrease in COX activity and ATP production is plausibly triggered by a set of mutations in the mitochondrial cytochrome *c* [67,68]. These mtDNA mutations appear to be compensated by restorer counter-adaptations within the population-specific coevolved nuclear backgrounds, which become ‘unmasked’ in F₂ hybrids [68]. Reduced fitness of interpopulation hybrids in this model is further correlated with elevated oxidative damage [69], and associated with differential expression of mitochondrial and nuclear OXPHOS genes [70], indicating the complexity of consequences triggered by the disruption of the coevolved mitonuclear lineages. Fitness of these hybrids can be restored via backcrossing to the maternal, not paternal, populations of origin, heavily implicating negative mitonuclear epistasis as the driver of the hybrid breakdown [71]. The impairment of fundamental functions—foremost the

expression of reproductive traits—in this model, highlights the critical role of mitonuclear interactions in upholding organismal fitness, and also the potential significance of these epistatic interactions to the evolution of reproductive isolation between incipient, allopatric populations [10,43,72].

Another model that has provided valuable insights into the role of interpopulation mitonuclear allelic interactions in organismal fitness, uses isonuclear fly lines of the fruit fly *D. melanogaster*, whereby distinct mitochondrial haplotypes sourced from different populations around the globe have been placed into a standardized foreign nuclear background [14,17]. Segregating nuclear allelic variation across these lines has been eliminated, such that modifications to the expression of phenotypes across the lines can be traced directly to variation in the mitochondrial genotype [17]. Fertility assessments among these lines suggest that all mitochondrial haplotypes confer lower male competitive fertility when expressed alongside an evolutionary novel nuclear background in comparison with the coevolved nuclear background [14]. Disruption of the coevolved mitonuclear genotype of one line rendered males, but not females, sterile when placed alongside one particular nuclear background [16,73]. These patterns of decreased fitness upon mitonuclear disruption are consistent with the hypothesis that coevolved nuclear genomes host compensatory counter-adaptations that offset deleterious mutations in the mtDNA. One study that harnessed these isonuclear lines reported that nuclear genome-wide gene expression patterns were influenced by cryptic polymorphisms within the mitochondrial genome, with these patterns virtually completely limited to males. Specifically, the expression of approximately 10% of all nuclear transcripts surveyed (approx. 1500) was sensitive to the mitochondrial haplotype in males, but only seven genes in total in females [73]. The stark male specificity of these mtDNA-induced effects is in line with the hypothesis that the maternal inheritance of the mitochondria will facilitate the build-up of mutations that are male-biased in their phenotypic effects under a sex-specific selective sieve, often referred to as *mother's curse* [15,73–76]. Even though the study did not explicitly examine mitonuclear interactions (i.e. the nuclear background was held isogenic), it revealed a candidate list of around 1500 nuclear genes whose expression is sensitive to interference by mutations harboured within the mitochondrial genome, but only in males. This list contains many genes whose expression is limited to the male reproductive tissues, and which serve essential roles in encoding male reproduction function, enriched for expression in the male reproductive tissues, and the gene list does not overlap considerably with the list of genes that are currently annotated as having mitochondrial-related function. Thus, these nuclear genes greatly increase the number of candidate nuclear genes for involvement in mitonuclear interactions, and extend the significance of the mitonuclear interaction well beyond that of simply being a regulator of core OXPHOS function [77].

Further compelling evidence for the profundity of mitonuclear interactions and their impact on key fitness traits comes from research in seed beetles [55,78] and yeast [79]. Arnqvist *et al.* [78] generated 25 fully crossed mitonuclear genotypes, via introgressive backcrossing of mtDNA haplotypes sourced from five distinct populations into the nuclear backgrounds associated with the same five populations, and subsequently measured the carbon dioxide production under two different temperature regimes. While no overall differences in metabolic performance between mitochondrial haplotypes

or nucleotypes were detected, complex gene-by-gene-by-environment interactions existed in the form of mitonuclear interactions whose outcomes were thermally sensitive. This finding fully aligns with the results of a similar experiment, harnessing the same 25 mitonuclear genotypes, which found that the development rates associated with particular mitonuclear genetic combinations were contingent on the temperature at which the beetles were reared [55]. That is, certain combinations of mitochondrial and nuclear genotype conferred relatively faster development time at the lower temperature, whereas other combinations had a relatively faster development time at the higher temperature. Generally, the superior mitonuclear genotypes at one temperature were not the superior performers at the other temperature. Interestingly, in four of five cases, the development times associated with co-evolved mitonuclear combinations was slower compared with those associated with disrupted mitonuclear gene complexes, counter to prediction.

Together, these studies demonstrate that mitonuclear interactions affect core fitness traits, but that the outcomes of these epistatic interactions depend on the prevailing environment. These findings fit the expectation that the mitonuclear interaction coevolves along population-specific trajectories, and towards population-specific optima in response to natural selection as a result of inhabiting distinct spatial and temporal environments. Considering the temperature sensitivity of OXPHOS enzyme activity, it is not surprising, in retrospect, that temperature has a strong effect on the efficiencies at which different combination of mitochondrial haplotype and nucleotype perform. The mix-and-matching that naturally occurs in panmictic populations, combined with the allelic variation that is known to persist within populations, will by default generate a plethora of distinct mitonuclear combinations, some of which will be most likely more efficient regulators of OXPHOS than others under certain environmental conditions.

While variation in mitochondrial genes, and thus in metabolic rate, is likely to play a pivotal role in the adaptability and evolvability of single populations to changing environments, mitochondrial incompatibilities will eventually invoke hybrid breakdown effects, ultimately marking the early stages of reproductive isolation between incipient populations. Because of this common pattern of negative mitonuclear epistasis in hybrids, and the role of mitochondrial function/dysfunction in organismal and reproductive fitness, mitochondrial bioenergetics has been hypothesized as a major driver of speciation [72]. Negative mitonuclear epistasis might act as an efficient barrier between species to reinforce reproductive isolation manifested as predicted by DMI [58–60]. Under this model, a deleterious mutation that reduces fitness can theoretically persist within allopatric populations as long as this mutation is rescued by a compensatory mutation at a second locus within respective populations. However, the compensatory effect may relinquish once breeding between individuals of these population occurs, leading to allelic mixtures that fail to combine the deleterious mutation with the compensatory allele, and conferring hybrid breakdown, at least of latter generation hybrids (F_2 and beyond). Considering the complexity of mitonuclear interaction, the distribution of mitochondrial components over two obligate genomes, and the different rates at which mitochondrial and nuclear loci evolve, the mitonuclear bond should be highly susceptible to DMI.

5. Interspecies interactions

Species-level evidence for mitonuclear mismatches must fundamentally be driven by evolutionary processes that are particular to populations. That is, the occurrence of macroevolutionary species-level mitonuclear incompatibilities must be the outcome of coevolutionary processes that take place within species and populations. As such, DMIs between allopatric populations are likely to progress through to speciation if isolation between populations persists over long enough evolutionary timescales to enable macroevolutionary patterns to manifest [31,60,61,80]. It is difficult to predict whether the degrees of mitonuclear mismatch, driven by DMI will increase as we move from the within species to between species levels. Intuitively, one might expect this increase to arise, given that the levels of genetic divergence increase markedly as one transitions through these scales.

Empirical evidence supporting the notion that mitonuclear incompatibilities increase with genetic distance comes from research using human cell lines stripped of mtDNA and then repopulated with the mtDNA from increasingly distantly related taxa [81,82]. A first set of experiments paired a human nucleotype with the mtDNA haplotype of the common chimpanzee, pigmy chimpanzee and gorilla [82]. Interruption of the coevolved mitonuclear lineage resulted in decreases in the activity of complex I of the mETS of around 40% in cybrids where the human nuclear background was paired with foreign mitochondrial genotypes in comparison with the native mtDNA genotype, whereas complexes II, III, IV and V were not affected [82]. In a second study, mitonuclear combinations were extended by combining the human cell line with mitochondrial haplotypes from even more distant taxa—orangutan, species representative of Old-World monkeys, New-World monkeys and lemurs—all of which failed to restore any tangible OXPHOS activity [81]. Similar mitonuclear incompatibilities have been documented in mtDNA-depleted cybrid cell lines of mouse species, repopulated with mtDNA from increasingly distant murine species [81–85]. By assessing respiratory capacities of single cell lines, these experiments reported varying decreases of activity for mETS complexes, with complex III most affected, displaying a striking decline in electron transport capacity with increasing species divergence [84]. Collectively, these experiments suggest that the disruption of the coevolved mitonuclear lineage leads to mitonuclear incompatibilities, reducing either OXPHOS function, complex assembly or increasing abnormally reacting oxygen species, and that the mismatch indeed increases with the level of taxonomic divergence in these groups.

The susceptibility of OXPHOS function to mitonuclear incompatibilities between species is perhaps best illustrated considering that all mitochondrial functions depend on nuclear factors that require import into the mitochondrion. These include mitochondrial transcription factors, transcription termination factors, RNA processing and modifying enzymes, ribosomal proteins, translation factors, all of which interact with and rely on specific sequence motifs in mitochondrial DNA or RNA to fulfil their functions [86], and which are expected to be sensitive to increasing genetic divergence between mitochondrial and nuclear genomes. Changes in recognition sites can thus readily disrupt mitochondrial translational machinery, leading to a breakdown in OXPHOS function. One such example comes from studies, which used

an *in vitro* system, combining factors of human and mouse mitochondrial transcription machinery, and found that mouse mitochondrial RNA polymerase (POLRMT) performs poorly when transcribing mtDNA using human promoters [87]. This work strongly suggests that the poor performance is linked to altered binding motifs of the transcription system establishing contact between POLRMT and mtDNA that have coevolved with their counterpart mtDNA recognition sites between the two species [70,87]. Another such example comes from a yeast hybrid model between *Saccharomyces cerevisiae* and *Saccharomyces bayanus* identifying the inability of the nuclear translation factor *AEP2* (*S. bayanus*) to regulate the translation of the mitochondrial F₀-ATP synthase subunit c (*OL11*, *S. cerevisiae*), causing sterility and sporulation defects [88,89]. Similar translational breakdown between species was observed between hybrids of *S. cerevisiae*, *S. bayanus* or *Saccharomyces paradoxus*, revealing the inability to properly splice mtDNA-encoded *COX1* via the interaction with the nuclear factor *Mrs1*, thus leading to sterility [89]. Similarly, recent hybridization studies in the fruit fly revealed a pronounced mitonuclear incompatibility, which was traced specifically to the mtDNA encoded tRNA^{TYR} of *Drosophila simulans* and the nuclear-encoded mitochondrial tyrosyl-tRNA synthetase of *D. melanogaster* [90]. This specific incompatibility decreases the activity of mETS complexes I, II and IV, compromises bristle formation, delays development and decreases fecundity [90]. A similar decline in OXPHOS capacity was previously documented in backcrosses of *D. simulans* and *Drosophila mauritiana* [91], and decreased fertility, fecundity and offspring viability, ultimately leading to hybrid breakdown in parasitoid wasps [92–94], and extensive mitonuclear epistatic interaction in the fruit fly [95].

Collectively, the empirical evidence outlined is compelling in its message that the phenotypic expression associated with specific mitochondrial genotypes is contingent upon the nuclear background alongside which these are expressed, and that these two genomes do not evolve along independent routes, but exhibit evolutionary trajectories that are tightly entwined with each other. Thus, the mitonuclear unit is a profound evolutionary unit, and mitonuclear coevolutionary processes are likely to impact on the general evolutionary dynamics of populations. What this means is that some mtDNA genotypes may perform well when coexpressed alongside certain nucleotypes, but may perform poorly alongside others, thereby significantly modulating key fitness traits. The reliance of physiological and reproductive traits on mitochondrial performance is nothing new, but what is unexpected is the extent to which coadaptive processes underpinning the mitonuclear interaction modify these traits. In hindsight, this may not be surprising, considering the different tempi and modes at which the interacting genomes evolve, and the perpetual process of coadaptation in response to selective pressure exerted by *de novo* mutations, standing genetic variation as well as distinct spatial and temporal environments. These epistatic interactions between mitochondrial and nuclear loci shape the adaptive landscapes and genetic architecture following population- and species-specific trajectories, and define mitonuclear complexes. The upshot of these processes is that nuclear and mitochondrial alleles of mitonuclear complexes are not freely interchangeable genetic elements, but rather act as a single, highly adapted and optimized functional unit. Efficient respiration is thus highly dependent on a good matching of mitochondrial and nuclear alleles, with suboptimal matches

invoking declines in fitness levels, because all fitness traits rely on mitochondrial function [77].

6. Intraindividual interactions and biomedical implications

The ubiquity of mitonuclear effects across multiple taxonomic scales suggests that mitonuclear interactions and incompatibilities will arise, and exert important consequences, wherever allelic variation in participating genes exists to fuel potential conflicts. We note that these conditions indeed exist at one further, but as yet unstudied, scale—within individuals. Intraindividual mitonuclear allelic variation can be fuelled by heteroplasmy in the mitochondrial genome, by heterozygosity and by the existence of gene duplicates (i.e. multiple varying copies of the same gene at different loci), and gene isoforms (i.e. splicing or length variants of the same gene) in the nuclear genome. What this means is that alternate mtDNA alleles can potentially be placed alongside alternate variants of nuclear-encoded mitochondrial genes, within one and the same individual. Assuming that all alleles are expressed, this gives rise to the possibility that mitochondrial complexes are assembled by a variety of different combinations of slightly varying subunits (protein isoforms), with the potential for enzymatic activities to be modified, exerting selective pressure on single complexes and mitonuclear combinations.

Support for this concept comes from two lines of research. First, mitonuclear allelic interactions have been observed in heteroplasmic individuals of fruit fly, where temperature-dependent selection of mitochondrial haplotypes, within individuals, has been shown to be contingent on the individual's coexpressed nuclear background [96]. Second, many species possess duplicates of nuclear-encoded mitochondrial genes, which exhibit tissue-specific (generally testis) expression. In one study on fruit flies, the expression of some mitochondrial gene duplicates was exclusively limited to testes and the expression of these duplicates outweighed the expression of the original parental genes threefold in the testes, whereas in all other tissues, gene duplicates were not expressed at all, instead fully relying on the expression of the parental variant [97].

An interesting case in point, which exemplifies the potential requirement of sex-specific functional variants of mitonuclear complexes, comes from the system of doubly uniparental transmission of mtDNA in some bivalves [98–100]. In this system, heteroplasmy is the norm: a maternal haplotype is transmitted to all offspring, and a paternal haplotype is exclusively transmitted to male offspring. In males, the maternal haplotype is predominant in all somatic tissues, but is outweighed by the paternal haplotype in the gonads, a system that may have evolved in response to asymmetries in functional requirements of reproductive tissues [101,102]. Interestingly, sequence divergence between the two haplotypes can exceed 40%, yet mitochondrial function is maintained, suggesting that mtDNA-encoded components may be matched alongside sex-specific isoforms or duplicates of nuclear-encoded mitochondrial components [101,102].

In mammals, the expression patterns of tissue-specific, other than testis-specific, variants is currently being elucidated. At least five complex IV protein subunits have tissue-specific isoforms. These are Cox4i1/Cox4i2; Cox6a1/

Cox6a2; Cox6b1/Cox6b2; Cox7a1/Cox7a2; Cox8a/Cox8b/Cox8c. Cox7a1, for example, is a heart/muscle isoform, whereas Cox7a2 is a liver isoform [103]. Moreover, expression patterns associated with particular isoforms, which confer different catalytic properties, do not only change across tissues, but also upon certain metabolic stimuli. Thus, hypoxia induced the replacement of the complex IV subunit 4–1 by its isoform 4–2, conferring a higher turnover rate to the activity of the enzyme [104]. A similar phenomenon was previously described in yeast mitochondria [105].

The presence of heteroplasmy in mtDNA, coupled with such context-dependence in expression of alternative mitochondrial gene isoforms, thus provides explicit scope for tissue-dependent intraindividual mitonuclear interactions, [97]. The existence of differential tissue-specific or physiologically induced expression of genes means that different combinations of mitochondrial components—with different catalytic properties—are expressed and assembled in different tissues or in response to particular stimuli. This componentry is labile and responsive to tissue-specific necessities, environmental changes or more generally to specific functional needs. In this respect, one must note that the variety of functions associated with the mETS is diverse and context-dependent, with different tissues requiring different, even contradictory, set-ups. For example, the main role of mETS in brown adipose tissue is to provide heat, whereas, in heart muscle, it is to provide ATP. However, maximizing ATP production efficiency implies reducing heat production. Less evident, but equally important, the physiological role of hepatocyte mETS is substantially different to that of neurons, to the extent that neurons cannot use fatty acids as a fuel because they lack β -oxidation [106].

Together, these observations strongly indicate that the within-individual level is not exempt from the effects of the mitonuclear interaction, because similar mechanisms, involving alternative variants of mitochondrial genes, exist within or across tissues in response to local or modified functional needs, or spontaneous mtDNA polymorphisms. Considering that heteroplasmy [11,40,44,45,107] and gene duplication of mitochondrial genes [46–48] are pervasive across taxa, the intraindividual level provides a new frontier for exploration of the evolutionary and medical significance of the mitonuclear interaction. A very striking example of the potential for intraindividual mitonuclear interaction was found in mice with artificially induced heteroplasmy between two wild-type mtDNAs, that of the Balb/cByJ mouse and that of the NZB/BINJ mouse [108]. It was robustly shown that selection on the mtDNA was tissue-specific, favouring NZB/BINJ mtDNA in the liver and kidney, but Balb/cByJ mtDNA in the blood and spleen, whereas other tissues showed no preference for either mtDNA type [108]. Differences in OXPHOS performance have been proposed to account for this phenomenon [21], which likely reflect the outcomes of mitonuclear interactions between the respective mtDNA types and different tissue-specific nuclear transcript isoforms.

Finally, we raise the prospect that mitonuclear interactions, at all scales from the intraindividual to between population levels, will not only be relevant to, but might help to explain much of the unaccounted for variation in the penetrance, variable array of symptoms and severity of human mitochondrial disease. The pervasiveness of the mitonuclear effects that we have outlined above, across numerous taxa and multiple biological scales, in itself suggests that humans will not be

exempt from the processes shaping mitonuclear coevolution and the ensuing effects. For instance, variance in mitonuclear compatibility could arise between divergent mitochondrial haplogroups and associated nucleotypes, which might predispose themselves to multifaceted and complex disease phenotypes [109]. It is well established that mitochondria are central to a multitude of disease phenotypes, yet, in many instances, straightforward links between mtDNA mutation and disease expression do not exist, and it remains largely unknown how this influence is exerted. We contend that in numerous instances mitochondrial disease may not be expressed through additive effects exerted by mitochondrial polymorphisms alone, but rather through epistasis between these polymorphisms and interacting nuclear alleles, and that this context-dependent expression of mitochondrial phenotypes may be key to understanding mitochondrial degenerative diseases. Particularly intriguing are those pathogenic mtDNA mutations that can reach homoplasmy and yet show very different penetrance among patients. It is puzzling that, with these homoplasmic mutations, some individuals suffer a severe life-threatening disease, whereas others are healthy [110]. It was proposed that functional epistasis between some nucleotypes and the mutant mtDNA may prevent disease [111].

Mitonuclear incompatibilities have previously been suggested to contribute to the expression of mitochondrial disease, and been associated with impairment of the mitochondrial translational machinery and OXPHOS assembly in LHON syndrome [112], Leigh syndrome [113], complex I-specific neurodegenerative disease [114] and in the phenotypic expression of deafness in humans and mice [20,115]. Heteroplasmy [44,45], gene duplication of nuclear-encoded mitochondrial genes [48,116], as well as the genetic footprint of adaptive selection in mitochondrial gene trees [117–119] are pervasive in humans, and all of these factors have the potential to fuel mitonuclear interactions for key phenotypes, as outlined above. Thus, the coevolutionary processes underpinning the mitonuclear interaction may provide tangible insights into our understanding of the origins of mitochondrial disease, and ultimately help us to predict incidences and severity of disease expression.

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References

- Gregersen N, Hansen J, Palmfeldt J. 2012 Mitochondrial proteomics—a tool for the study of metabolic disorders. *J. Inherit. Metab. Dis.* **35**, 715–726. (doi:10.1007/s10545-012-9480-3)
- Sickmann A *et al.* 2003 The proteome of *Saccharomyces cerevisiae* mitochondria. *Proc. Natl Acad. Sci. USA* **100**, 13 207–13 212. (doi:10.1073/pnas.2135385100)
- Mootha VK *et al.* 2003 Integrated analysis of protein composition, tissue diversity, and gene regulation in mouse mitochondria. *Cell* **115**, 629–640. (doi:10.1016/S0092-8674(03)00926-7)
- Lotz C *et al.* 2013 The characterization, design, and function of the mitochondrial proteome: from organs to organisms. *J. Proteome Res.* **13**, 433–446. (doi:10.1021/pr400539j)
- Ryan MT, Hoogenraad NJ. 2007 Mitochondrial-nuclear communications. *Annu. Rev. Biochem.* **76**, 701–722. (doi:10.1146/annurev.biochem.76.052305.091720)
- McKenzie M, Lazarou M, Thorburn DR, Ryan MT. 2007 Analysis of mitochondrial subunit assembly into respiratory chain complexes using blue native polyacrylamide gel electrophoresis. *Anal. Biochem.* **364**, 128–137. (doi:10.1016/j.ab.2007.02.022)
- Wolstenholme DR. 1992 Animal mitochondrial DNA: structure and evolution. In *Mitochondrial genomes* (eds DR Wolstenholme, KW Jeon), pp. 173–213. Maryland Heights, MO: Elsevier.
- Wallace DC. 2010 Mitochondrial DNA mutations in disease and aging. *Environ. Mol. Mutagen.* **51**, 440–450. (doi:10.1002/em.20586)
- Blier PU, Dufresne F, Burton RS. 2001 Natural selection and the evolution of mtDNA-encoded peptides: evidence for intergenomic co-adaptation. *Trends Genet.* **17**, 400–406. (doi:10.1016/S0168-9525(01)02338-1)
- Dowling DK, Friberg U, Lindell J. 2008 Evolutionary implications of non-neutral mitochondrial genetic variation. *Trends Ecol. Evol.* **23**, 546–554. (doi:10.1016/j.tree.2008.05.011)
- Rand DM. 2001 The units of selection on mitochondrial DNA. *Annu. Rev. Ecol. Syst.* **32**, 415–448. (doi:10.1146/annurev.ecolsys.32.081501.114109)
- Ballard JW, Whitlock MC. 2004 The incomplete natural history of mitochondria. *Mol. Ecol.* **13**, 729–744. (doi:10.1046/j.1365-294X.2003.02063)
- Galtier N, Nabholz B, Glémin S, Hurst GDD. 2009 Mitochondrial DNA as a marker of molecular diversity: a reappraisal. *Mol. Ecol.* **18**, 4541–4550. (doi:10.1111/j.1365-294X.2009.04380.x)
- Yee WK, Sutton KL, Dowling DK. 2013 *In vivo* male fertility is affected by naturally occurring mitochondrial haplotypes. *Curr. Biol.* **23**, R55–R56. (doi:10.1016/j.cub.2012.12.002)
- Camus MF, Clancy DJ, Dowling DK. 2012 Mitochondria, maternal inheritance, and male aging. *Curr. Biol.* **22**, 1–5. (doi:10.1016/j.cub.2012.07.018)
- Clancy DJ, Hime GR, Shirras AD. 2011 Cytoplasmic male sterility in *Drosophila melanogaster* associated with a mitochondrial CYTB variant. *Heredity* **107**, 374–376. (doi:10.1038/hdy.2011.12)
- Clancy DJ. 2008 Variation in mitochondrial genotype has substantial lifespan effects which may be modulated by nuclear background. *Aging Cell.* **7**, 795–804. (doi:10.1111/j.1474-9726.2008.00428.x)
- Ruiz-Pesini E *et al.* 2000 Human mtDNA haplogroups associated with high or reduced spermatozoa motility. *Am. J. Hum. Genet.* **67**, 682–696. (doi:10.1086/303040)
- Montiel-Sosa F, Ruiz-Pesini E, Enriquez JA, Marcuello A, Diez-Sanchez C, Montoya J, Wallace DC, Lopez-Perez MJ. 2006 Differences of sperm motility in mitochondrial DNA haplogroup U sublineages. *Gene* **368**, 21–27. (doi:10.1016/j.gene.2005.09.015)
- Johnson KR, Zheng QY, Bykhovskaya Y, Spirina O, Fischel-Ghodsian N. 2001 A nuclear-mitochondrial DNA interaction affecting hearing impairment in mice. *Nat. Genet.* **27**, 191–194. (doi:10.1038/84831)
- Moreno-Loshuertos R, Acin-Perez R, Fernandez-Silva P, Movilla N, Perez-Martos A, Rodriguez de Cordoba S, Gallardo ME, Enriquez JA. 2006 Differences in reactive oxygen species production explain the phenotypes associated with common mouse mitochondrial DNA variants. *Nat. Genet.* **38**, 1261–1268. (doi:10.1038/ng1897)
- Roubertoux PL *et al.* 2003 Mitochondrial DNA modifies cognition in interaction with the nuclear genome and age in mice. *Nat. Genet.* **35**, 65–69. (doi:10.1038/ng1230)
- MITOMAP: a human mitochondrial genome database.* See <http://www.mitomap.org>.
- Goto Y, Nonaka I, Horai S. 1990 A mutation in the tRNA(Leu)(UUR) gene associated with the MELAS subgroup of mitochondrial encephalomyopathies. *Nature* **348**, 651–653. (doi:10.1038/348651a0)
- Wallace D, Singh G, Lott M, Hodge J, Schurr T, Lezza A, Elsas L, Nikoskelainen E. 1988 Mitochondrial DNA mutation associated with Leber's hereditary optic neuropathy. *Science* **242**, 1427–1430. (doi:10.1126/science.3201231)

26. Patti ME, Corvera S. 2010 The role of mitochondria in the pathogenesis of type 2 diabetes. *Endocr. Rev.* **31**, 364–395. (doi:10.1210/er.2009-0027)
27. Piaceri I, Rinnoci V, Bagnoli S, Failli Y, Sorbi S. 2012 Mitochondria and Alzheimer's disease. *J. Neurol. Sci.* **322**, 31–34. (doi:10.1016/j.jns.2012.05.033)
28. Mizuno Y, Ikebe S, Hattori N, Nakagawa-Hattori Y, Mochizuki H, Tanaka M, Ozawa T. 1995 Role of mitochondria in the etiology and pathogenesis of Parkinson's disease. *Biochim. Biophys. Acta* **1271**, 265–274. (doi:10.1016/0925-4439(95)00038-6)
29. Schon EA, DiMauro S, Hirano M. 2012 Human mitochondrial DNA: roles of inherited and somatic mutations. *Nat. Rev. Genet.* **13**, 878–890. (doi:10.1038/nrg3275)
30. Wallace DC. 2012 Mitochondria and cancer. *Nat. Rev. Cancer* **12**, 685–698. (doi:10.1038/nrc3365)
31. Grossman LI, Shoubridge EA. 1996 Mitochondrial genetics and human disease. *Bioessays* **18**, 983–991. (doi:10.1002/bies.950181208)
32. Wallace DC. 2005 A mitochondrial paradigm of metabolic and degenerative diseases, aging, and cancer: a dawn for evolutionary medicine. *Annu. Rev. Genet.* **39**, 359–407. (doi:10.1146/annurev.genet.39.110304.095751)
33. Wallace DC. 1992 Diseases of the mitochondrial DNA. *Annu. Rev. Biochem.* **61**, 1175–1212. (doi:10.1146/annurev.bi.61.070192.005523)
34. Rand DM, Haney RA, Fry AJ. 2004 Cytonuclear coevolution: the genomics of cooperation. *Trends Ecol. Evol.* **19**, 645–653. (doi:10.1016/j.tree.2004.10.003)
35. Lynch M, Koskella B, Schaack S. 2006 Mutation pressure and the evolution of organelle genomic architecture. *Science* **311**, 1727–1730. (doi:10.1126/science.1118884)
36. Lynch M. 1996 Mutation accumulation in transfer RNAs: molecular evidence for Muller's ratchet in mitochondrial genomes. *Mol. Biol. Evol.* **13**, 209–220. (doi:10.1093/oxfordjournals.molbev.a025557)
37. Hagstrom E, Freyer C, Battersby BJ, Stewart JB, Larsson NG. 2013 No recombination of mtDNA after heteroplasmy for 50 generations in the mouse maternal germline. *Nucleic Acids Res.* **42**, 1111–1116. (doi:10.1093/nar/gkt969)
38. Piganeau G, Gardner M, Eyre-Walker A. 2004 A broad survey of recombination in animal mitochondria. *Mol. Biol. Evol.* **21**, 2319–2325. (doi:10.1093/molbev/msh244)
39. Tsaousis AD, Martin DP, Ladoukakis ED, Posada D, Zouros E. 2005 Widespread recombination in published animal mtDNA sequences. *Mol. Biol. Evol.* **22**, 925–933. (doi:10.1093/molbev/msi084)
40. White DJ, Wolff JN, Pierson M, Gemmell NJ. 2008 Revealing the hidden complexities of mtDNA inheritance. *Mol. Ecol.* **17**, 4925–4942. (doi:10.1111/j.1365-294X.2008.03982.x)
41. Lynch M. 1997 Mutation accumulation in nuclear, organelle, and prokaryotic transfer RNA genes. *Mol. Biol. Evol.* **14**, 914–925. (doi:10.1093/oxfordjournals.molbev.a025834)
42. Osada N, Akashi H. 2012 Mitochondrial–nuclear interactions and accelerated compensatory evolution: evidence from the primate cytochrome c oxidase complex. *Mol. Biol. Evol.* **29**, 337–346. (doi:10.1093/molbev/msr211)
43. Barreto FS, Burton RS. 2013 Evidence for compensatory evolution of ribosomal proteins in response to rapid divergence of mitochondrial rRNA. *Mol. Biol. Evol.* **30**, 310–314. (doi:10.1093/molbev/mss228)
44. Payne BAI *et al.* 2013 Universal heteroplasmy of human mitochondrial DNA. *Hum. Mol. Genet.* **22**, 384–390. (doi:10.1093/hmg/dd5435)
45. Avital G, Buchshtav M, Zhidkov I, Tuval Feder J, Dadon S, Rubin E, Glass D, Spector TD, Mishmar D. 2012 Mitochondrial DNA heteroplasmy in diabetes and normal adults: role of acquired and inherited mutational patterns in twins. *Hum. Mol. Genet.* **21**, 4214–4224. (doi:10.1093/hmg/dd5245)
46. Bai Y, Casola C, Feschotte C, Betran E. 2007 Comparative genomics reveals a constant rate of origination and convergent acquisition of functional retrogenes in *Drosophila*. *Genome Biol.* **8**, R11. (doi:10.1186/gb-2007-8-1-r11)
47. Tripoli G, D'Elia D, Barsanti P, Caggese C. 2005 Comparison of the oxidative phosphorylation (OXPHOS) nuclear genes in the genomes of *Drosophila melanogaster*, *Drosophila pseudoobscura* and *Anopheles gambiae*. *Genome Biol.* **6**, R11. (doi:10.1186/gb-2005-6-2-r11)
48. Emerson JJ, Kaessmann H, Betran E, Long M. 2004 Extensive gene traffic on the mammalian X chromosome. *Science* **303**, 537–540. (doi:10.1126/science.1090042)
49. Gregorius HR, Ross MD. 1984 Selection with gene-cytoplasm interactions. *Genetics* **107**, 165.
50. Rand DM, Clark AG, Kann LM. 2001 Sexually antagonistic cytonuclear fitness interactions in *Drosophila melanogaster*. *Genetics* **159**, 173–187.
51. Clark AG. 1984 Natural selection with nuclear and cytoplasmic transmission. *Genetics* **107**, 679.
52. Babcock CS, Asmussen MA. 1996 Effects of differential selection in the sexes on cytonuclear polymorphism and disequilibria. *Genetics* **144**, 839–853.
53. Babcock CS, Asmussen MA. 1998 Effects of differential selection in the sexes on cytonuclear dynamics. Life stages with sex differences. *Genetics* **149**, 2063–2077.
54. Dowling DK, Friberg U, Hailer F, Arnqvist G. 2007 Intergenomic epistasis for fitness: within-population interactions between cytoplasmic and nuclear genes in *Drosophila melanogaster*. *Genetics* **175**, 235–244. (doi:10.1534/genetics.105.052050)
55. Dowling DK, Abiega KC, Arnqvist G. 2007 Temperature-specific outcomes of cytoplasmic–nuclear interactions on egg-to-adult development time in seed beetles. *Evolution* **61**, 194–201. (doi:10.1111/j.1558-5646.2007.00016.x)
56. Clark AG, Lyckegaard EM. 1988 Natural selection with nuclear and cytoplasmic transmission. *Genetics* **118**, 471.
57. Maklakov AA, Friberg U, Dowling DK, Arnqvist G. 2006 Within-population variation in cytoplasmic genes affects female life span and aging in *Drosophila melanogaster*. *Evolution* **60**, 2081–2086. (doi:10.1111/j.0014-3820.2006.tb01845.x)
58. Muller HJ. 1942 Isolating mechanisms, evolution, and temperature. *Biol. Symp.* **6**, 71–125.
59. Dobzhansky T. 1936 Studies on hybrid sterility. II. Localization of sterility factors in *Drosophila pseudoobscura* hybrids. *Genetics* **21**, 113–135.
60. Orr HA. 1995 The population genetics of speciation: the evolution of hybrid incompatibilities. *Genetics* **139**, 1805–1813.
61. Burton RS, Barreto FS. 2012 A disproportionate role for mtDNA in Dobzhansky–Muller incompatibilities? *Mol. Ecol.* **21**, 4942–4957. (doi:10.1111/mec.12006)
62. Burton RS. 1986 Evolutionary consequences of restricted gene flow among natural populations of the copepod, *Triopiopus californicus*. *Bull. Mar. Sci.* **39**, 526–535.
63. Burton RS. 1990 Hybrid breakdown in developmental time in the copepod *Tigriopus californicus*. *Evolution* **44**, 1814–1822. (doi:10.2307/2409510)
64. Edmands S. 1999 Heterosis and outbreeding depression in interpopulation crosses spanning a wide range of divergence. *Evolution* **53**, 1757–1768. (doi:10.2307/2640438)
65. Rawson PD, Burton RS. 2002 Functional coadaptation between cytochrome c and cytochrome c oxidase within allopatric populations of a marine copepod. *Proc. Natl Acad. Sci. USA* **99**, 12 955–12 958. (doi:10.1073/pnas.202335899)
66. Ellison CK, Burton RS. 2006 Disruption of mitochondrial function in interpopulation hybrids of *Tigriopus californicus*. *Evolution* **60**, 1382–1391. (doi:10.1111/j.0014-3820.2006.tb01217.x)
67. Rawson PD, Brazeau DA, Burton RS. 2000 Isolation and characterization of cytochrome c from the marine copepod *Tigriopus californicus*. *Gene* **248**, 15–22. (doi:10.1016/S0378-1119(00)00145-1)
68. Harrison JS, Burton RS. 2006 Tracing hybrid incompatibilities to single amino acid substitutions. *Mol. Biol. Evol.* **23**, 559–564. (doi:10.1093/molbev/msj058)
69. Barreto FS, Burton RS. 2013 Elevated oxidative damage is correlated with reduced fitness in interpopulation hybrids of a marine copepod. *Proc. R. Soc. B* **280**, 20131521. (doi:10.1098/rspb.2013.1521)
70. Ellison CK, Burton RS. 2008 Genotype-dependent variation of mitochondrial transcriptional profiles in interpopulation hybrids. *Proc. Natl Acad. Sci. USA* **105**, 15 831–15 836. (doi:10.1073/pnas.0804253105)
71. Ellison CK, Burton RS. 2008 Interpopulation hybrid breakdown maps to the mitochondrial genome. *Evolution* **62**, 631–638. (doi:10.1111/j.1558-5646.2007.00305.x)
72. Gershoni M, Templeton AR, Mishmar D. 2009 Mitochondrial bioenergetics as a major motive force of speciation. *Bioessays* **31**, 642–650. (doi:10.1002/bies.200800139)
73. Innocenti P, Morrow EH, Dowling DK. 2011 Experimental evidence supports a sex-specific

- selective sieve in mitochondrial genome evolution. *Science* **332**, 845–848. (doi:10.1126/science.1201157)
74. Frank SA, Hurst LD. 1996 Mitochondria and male disease. *Nature* **383**, 224. (doi:10.1038/383224a0)
75. Gemmell NJ, Metcalf VJ, Allendorf FW. 2004 Mother's curse: the effect of mtDNA on individual fitness and population viability. *Trends Ecol. Evol.* **19**, 238–244. (doi:10.1016/j.tree.2004.02.002)
76. Wolff JN, Gemmell NJ. 2013 Mitochondria, maternal inheritance, and asymmetric fitness: why males die younger. *Bioessays* **35**, 93–99. (doi:10.1002/bies.201200141)
77. Lane N. 2011 Mitonuclear match: optimizing fitness and fertility over generations drives ageing within generations. *Bioessays* **33**, 860–869. (doi:10.1002/bies.201100051)
78. Arnqvist G, Dowling DK, Eady P, Gay L, Tregenza T, Tuda M, Hosken DJ. 2010 Genetic architecture of metanolic rate: environment specific epistasis between mitochondrial and nuclear genes in an insect. *Evolution* **64**, 3354–3363. (doi:10.1111/j.1558-5646.2010.01135.x)
79. Zeyl C, Andreson B, Weninck E. 2005 Nuclear-mitochondrial epistasis for fitness in *Saccharomyces cerevisiae*. *Evolution* **59**, 910–914. (doi:10.1111/j.0014-3820.2005.tb01764.x)
80. Oliveira DC, Raychoudhury R, Lavrov DV, Werren JH. 2008 Rapidly evolving mitochondrial genome and directional selection in mitochondrial genes in the parasitic wasp *Nasonia* (Hymenoptera: Pteromalidae). *Mol. Biol. Evol.* **25**, 2167–2180. (doi:10.1093/molbev/msn159)
81. Kenyon L, Moraes CT. 1997 Expanding the functional human mitochondrial DNA database by the establishment of primate xenomitochondrial cybrids. *Proc. Natl Acad. Sci. USA* **94**, 9131–9135. (doi:10.1073/pnas.94.17.9131)
82. Barrientos A, Kenyon L, Moraes CT. 1998 Human xenomitochondrial cybrids. Cellular models of mitochondrial complex I deficiency. *J. Biol. Chem.* **273**, 14 210–14 217. (doi:10.1074/jbc.273.23.14210)
83. Yamaoka M, Isobe K, Shitara H, Yonekawa H, Miyabayashi S, Hayashi JI. 2000 Complete repopulation of mouse mitochondrial DNA-less cells with rat mitochondrial DNA restores mitochondrial translation but not mitochondrial respiratory function. *Genetics* **155**, 301–307.
84. McKenzie M, Chiotis M, Pinkert CA, Trounce IA. 2003 Functional respiratory chain analyses in murine xenomitochondrial cybrids expose coevolutionary constraints of cytochrome b and nuclear subunits of complex III. *Mol. Biol. Evol.* **20**, 1117–1124. (doi:10.1093/molbev/msg132)
85. Dey R, Barrientos A, Moraes CT. 2000 Functional constraints of nuclear–mitochondrial DNA interactions in xenomitochondrial rodent cell lines. *J. Biol. Chem.* **275**, 31 520–31 527. (doi:10.1074/jbc.M004053200)
86. Smits P, Smeitink J, van den Heuvel L. 2010 Mitochondrial translation and beyond: processes implicated in combined oxidative phosphorylation deficiencies. *J. Biomed. Biotechnol.* **2010**, 737385. (doi:10.1155/2010/737385)
87. Gaspari M, Falkenberg M, Larsson NG, Gustafsson CM. 2004 The mitochondrial RNA polymerase contributes critically to promoter specificity in mammalian cells. *EMBO J.* **23**, 4606–4614. (doi:10.1038/sj.emboj.7600465)
88. Lee HY, Chou JY, Cheong L, Chang NH, Yang SY, Leu JY. 2008 Incompatibility of nuclear and mitochondrial genomes causes hybrid sterility between two yeast species. *Cell* **135**, 1065–1073. (doi:10.1016/j.cell.2008.10.047)
89. Chou JY, Hung YS, Lin KH, Lee HY, Leu JY. 2010 Multiple molecular mechanisms cause reproductive isolation between three yeast species. *PLoS Biol.* **8**, e1000432. (doi:10.1371/journal.pbio.1000432)
90. Meiklejohn CD, Holmbeck MA, Siddiq MA, Abt DN, Rand DM, Montooth KL. 2013 An incompatibility between a mitochondrial tRNA and its nuclear-encoded tRNA synthetase compromises development and fitness in *Drosophila*. *PLoS Genet.* **9**, e1003238. (doi:10.1371/journal.pgen.1003238)
91. Sackton TB, Haney RA, Rand DM. 2003 Cytonuclear coadaptation in *Drosophila*: disruption of cytochrome c oxidase activity in backcross genotypes. *Evolution* **57**, 2315–2325. (doi:10.1111/j.0014-3820.2003.tb00243.x)
92. Ellison CK, Niehuis O, Gadau J. 2008 Hybrid breakdown and mitochondrial dysfunction in hybrids of *Nasonia* parasitoid wasps. *J. Evol. Biol.* **21**, 1844–1851. (doi:10.1111/j.1420-9101.2008.01608.x)
93. Niehuis O, Judson AK, Gadau J. 2008 Cytonuclear genic incompatibilities cause increased mortality in male F2 hybrids of *Nasonia giraulti* and *N. vitripennis*. *Genetics* **178**, 413–426. (doi:10.1534/genetics.107.080523)
94. Koevoets T, Niehuis O, van de Zande L, Beukeboom LW. 2012 Hybrid incompatibilities in the parasitic wasp genus *Nasonia*: negative effects of hemizygoty and the identification of transmission ratio distortion loci. *Heredity* **108**, 302–311. (doi:10.1038/hdy.2011.75)
95. Rand DM, Fry A, Sheldahl L. 2006 Nuclear-mitochondrial epistasis and drosophila aging: introgression of *Drosophila simulans* mtDNA modifies longevity in *D. melanogaster* nuclear backgrounds. *Genetics* **172**, 329–341. (doi:10.1534/genetics.105.046698)
96. Doi A, Suzuki H, Matsuura ET. 1999 Genetic analysis of temperature-dependent transmission of mitochondrial DNA in *Drosophila*. *Heredity* **82**, 555–560. (doi:10.1038/sj.hdy.6885080)
97. Gallach M, Chandrasekaran C, Betran E. 2010 Analyses of nuclearly encoded mitochondrial genes suggest gene duplication as a mechanism for resolving intralocus sexually antagonistic conflict in *Drosophila*. *Genome Biol. Evol.* **2**, 835–850. (doi:10.1093/gbe/evq069)
98. Theologidis I, Saavedra C, Zouros E. 2007 No evidence for absence of paternal mtDNA in male progeny from pair matings of the mussel *Mytilus galloprovincialis*. *Genetics* **176**, 1367–1369. (doi:10.1534/genetics.106.069930)
99. Zouros E, Oberhauser Ball A, Saavedra C, Freeman KR. 1994 An unusual type of mitochondrial DNA inheritance in the blue mussel *Mytilus*. *Proc. Natl Acad. Sci. USA* **91**, 7463–7467. (doi:10.1073/pnas.91.16.7463)
100. Breton S, Beaupre HD, Stewart DT, Hoeh WR, Blier PU. 2007 The unusual system of doubly uniparental inheritance of mtDNA: isn't one enough? *Trends Genet.* **23**, 465–474. (doi:10.1016/j.tig.2007.05.011)
101. Doucet-Beaupre H, Breton S, Chapman EG, Blier PU, Bogan AE, Stewart DT, Hoeh WR. 2010 Mitochondrial phylogenomics of the Bivalvia (Mollusca): searching for the origin and mitogenomic correlates of doubly uniparental inheritance of mtDNA. *BMC Evol. Biol.* **10**, 50. (doi:10.1186/1471-2148-10-50)
102. Obata M, Sano N, Komaru A. 2011 Different transcriptional ratios of male and female transmitted mitochondrial DNA and tissue-specific expression patterns in the blue mussel, *Mytilus galloprovincialis*. *Dev. Growth Differ.* **53**, 878–886. (doi:10.1111/j.1440-169X.2011.01294.x)
103. Fornuskova D, Stiburek L, Wenchich L, Vinsova K, Hansikova H, Zeman J. 2010 Novel insights into the assembly and function of human nuclear-encoded cytochrome c oxidase subunits 4, 5a, 6a, 7a and 7b. *Biochem. J.* **428**, 363–374. (doi:10.1042/BJ20091714)
104. Fukuda R, Zhang H, Kim JW, Shimoda L, Dang CV, Semenza GL. 2007 HIF-1 regulates cytochrome oxidase subunits to optimize efficiency of respiration in hypoxic cells. *Cell* **129**, 111–122. (doi:10.1016/j.cell.2007.01.047)
105. Waterland RA, Basu A, Chance B, Poyton RO. 1991 The isoforms of yeast cytochrome c oxidase subunit V alter the *in vivo* kinetic properties of the holoenzyme. *J. Biol. Chem.* **266**, 4180–4186.
106. Speijer D. 2011 Oxygen radicals shaping evolution: why fatty acid catabolism leads to peroxisomes while neurons do without it. *Bioessays* **33**, 88–94. (doi:10.1002/bies.201000097)
107. Wolff JN, Nafisinia M, Sutovsky P, Ballard JWO. 2012 Paternal transmission of mitochondrial DNA as an integral part of mitochondrial inheritance in metapopulations of *Drosophila simulans*. *Heredity* **10**, 57–62. (doi:10.1038/hdy.2012.60)
108. Jenuth JP, Peterson AC, Shoubridge EA. 1997 Tissue-specific selection for different mtDNA genotypes in heteroplasmic mice. *Nat. Genet.* **16**, 93–95. (doi:10.1038/ng0597-93)
109. Reinhardt K, Dowling DK, Morrow EH. 2013 Mitochondrial replacement, evolution, and the clinic. *Science* **341**, 1345–1346. (doi:10.1126/science.1237146)
110. Limongelli A, Schaefer J, Jackson S, Invernizzi F, Kirino Y, Suzuki T, Reichmann H, Zeviani M. 2004 Variable penetrance of a familial progressive necrotising encephalopathy due to a novel tRNA(Ile) homoplasmic mutation in the mitochondrial genome. *J. Med. Genet.* **41**, 342–349. (doi:10.1136/jmg.2003.016048)
111. Moreno-Loshuertos R, Ferrin G, Acin-Perez R, Gallardo ME, Viscomi C, Perez-Martos A, Zeviani M,

- Fernandez-Silva P, Enriquez JA. 2011 Evolution meets disease: penetrance and functional epistasis of mitochondrial tRNA mutations. *PLoS Genet.* **7**, e1001379. (doi:10.1371/journal.pgen.1001379)
112. Pello R *et al.* 2008 Mitochondrial DNA background modulates the assembly kinetics of OXPHOS complexes in a cellular model of mitochondrial disease. *Hum. Mol. Genet.* **17**, 4001–4111. (doi:10.1093/hmg/ddn303)
113. Hoefs SJ *et al.* 2009 Baculovirus complementation restores a novel NDUFA2 mutation causing complex I deficiency. *Hum. Mutat.* **30**, E728–E736. (doi:10.1002/humu.21037)
114. Potluri P *et al.* 2009 A novel NDUFA1 mutation leads to a progressive mitochondrial complex I-specific neurodegenerative disease. *Mol. Genet. Metab.* **96**, 189–195. (doi:10.1016/j.ymgme.2008.12.004)
115. Guan MX *et al.* 2006 Mutation in TRMU related to transfer RNA modification modulates the phenotypic expression of the deafness-associated mitochondrial 12S ribosomal RNA mutations. *Am. J. Hum. Genet.* **79**, 291–302. (doi:10.1086/506389)
116. Vinckenbosch N, Dupanloup I, Kaessmann H. 2006 Evolutionary fate of retroposed gene copies in the human genome. *Proc. Natl Acad. Sci. USA* **103**, 3220–3225. (doi:10.1073/pnas.0511307103)
117. Ruiz-Pesini E, Wallace DC. 2006 Evidence for adaptive selection acting on the tRNA and rRNA genes of human mitochondrial DNA. *Hum. Mutat.* **27**, 1072–1081. (doi:10.1002/humu.20378)
118. Ruiz-Pesini E, Mishmar D, Brandon M, Procaccio V, Wallace DC. 2004 Effects of purifying and adaptive selection on regional variation in human mtDNA. *Science* **303**, 223–226. (doi:10.1126/science.1088434)
119. Mishmar D *et al.* 2003 Natural selection shaped regional mtDNA variation in humans. *Proc. Natl Acad. Sci. USA* **100**, 171–176. (doi:10.1073/pnas.0136972100)