Inception of a false memory by optogenetic manipulation of a hippocampal memory engram

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Memories can be easily distorted, and a lack of relevant animal models has largely hindered our understanding of false-memory formation. Here, we first identified a population of cells in the dentate gyrus (DG) of the hippocampus that bear the engrams for a specific context; these cells were naturally activated during the encoding phase of fear conditioning and their artificial reactivation using optogenetics in an unrelated context was sufficient for inducing the fear memory specific to the conditioned context. In a further study, DG or CA1 neurons activated by exposure to a particular context were labelled with channelrhodopsin-2 (ChR2). These neurons were later optically reactivated during fear conditioning in a different context. The DG experimental group showed increased freezing in the original context in which a foot shock was never delivered. The recall of this false memory was context specific, activated similar downstream regions engaged during natural fear-memory recall, and was also capable of driving an active fear response. Together, our data demonstrate that by substituting a natural conditioned stimulus with optogenetically reactivated DG cells that bear contextual memory engrams, it is possible to incept an internally and behaviourally represented false fear memory.

1. Introduction

Hebb’s [1] pioneering conceptualization of synaptic plasticity in 1949 followed by Bliss & Lomo’s [2] discovery of long-term potentiation (LTP) has provided the principal framework with which neuroscientists have pursued the neural mechanisms subserving learning and memory. Additional mechanisms, including alterations in membrane excitability and activation of immediately early genes (IEGs) at the whole single-cell level, as well as biochemical and structural alterations of dendritic spines have been recognized [3]. In neuropsychology, Semon [4] put forward the ‘engram’ theory of memory in the early twentieth century, which in current terms could be roughly stated as: when a memory is formed, a subpopulation of neurons will be excited and stay excited latently for the storage of the memory information (engram). When part of the total information at the time of storage is subsequently available, it will re-excite the engram for recall. Figure 1 presents a conceptual diagram of an engram-bearing neuronal population that incorporates neurobiological mechanisms including LTP.

A recent study showed that a selective post-training ablation of an IEG (i.e. CREB)-rich cell population in the amygdala that was activated during a fear-memory task results in a loss of that fear memory [5]. These data are consistent with the engram theory, but the final test of any hypothesis concerning memory engrams must be a mimicry experiment, in which apparent memory is generated (expressed) artificially without the usual requirement for sensory experience [6]. We recently reported such an experiment. In this article, we first provide a brief summary of this work. As the main body of this article, we then proceed to our more recent work on a mouse model of false memory that was made possible by the identification of memory engram-bearing cells.

We routinely use memories as guides for cognition and behaviour [6–9]. Memory, however, can often be unreliable because it is not a carbon copy...
2. Identification of contextual memory engrams in the dentate gyrus (see [16] for complete text and methods)

To label and reactivate a subpopulation of dentate gyrus (DG) cells active during the encoding of a memory, we targeted the DG of c-fos–tTA transgenic mice [17] with a TRE-ChR2-EYFP virus and an optical fibre implant (figure 2a). This approach couples the promoter of c-fos, an IEG often used as a marker of recent neuronal activity [18,19], to the tetracycline transactivator (tTA), a key component of the doxycycline (Dox) system for inducible expression of a gene of interest [20]. In our system, the absence of Dox permits c-fos-promoter-driven tTA to bind to its target tetracycline-responsive element (TRE) site, which in turn drives channelrhodopsin-2 (ChR2)–enhanced yellow fluorescent protein (EYFP) expression in neurons active during this defined period. These neurons can then be reactivated by light stimulation during testing (figure 2b,c). In the presence of Dox, however, tTA is blocked from binding to TRE and neurons active during this Dox-on period remains unlabelled by ChR2–EYFP.

We first tested whether the reactivation of a population of DG neurons active during the encoding phase of a fear memory was sufficient for the reinstatement of that memory. The experimental group (Exp) consisted of c-fos–tTA mice injected with TRE-ChR2-EYFP and implanted with an optical fibre targeting the dorsal DG (figure 2a). Mice were kept on Dox and underwent a habituation period to record their basal level of freezing in one context (context A) during which they received both light-off and light-on epochs. Next, they were taken off Dox and underwent fear conditioning in a distinct chamber (context B) in which a tone was paired with shock. The mice were then subjected to testing sessions with light-off and light-on epochs in context A while being back on Dox to prevent any subsequent labelling of active DG cells (figure 2c–g). During the habituation sessions, the Exp mice showed very little freezing during either light-off or light-on epochs. By contrast, after fear conditioning—during which putative fear memory engram-bearing DG cells were labelled with ChR2–EYFP—freezing levels during light-on epochs were higher compared with light-off epochs, which indicated light-induced fear-memory recall (figure 3a). A group of mice (NS group) that went through exactly the same procedures as the experimental group except that no shock was delivered during the training session did not freeze above background levels when the light was shone during the post-training session (figure 3b). Another group of mice (EYFP group) that went through the same experimental protocol except that the virus had no ChR2 gene also...
showed no augmented post-training freezing upon the light delivery (figure 3c). On the other hand, the post-training freezing levels were increased even more than those of the experimental group when the Dox-off training period was reduced to 1 day (Exp-1 day group, figure 3d), presumably because non-specific environmental stimuli, which compete with the specific context stimuli, were reduced. The level of freezing was increased further when the manipulation was performed bilaterally (Exp-Bi group, figure 3e).

The overall results (figure 3f) suggest that DG cells that express endogenous c-Fos during training, and therefore become labelled by ChR2–EYFP, define an active neural population that is sufficient for memory recall upon subsequent reactivation [16].

3. Inception of a false memory (see [21] for complete text and methods)

Having identified contextual engram-bearing cells in the DG, we went on to test whether an artificial conditioned (CS)–unconditioned stimulus (US) association—what we refer to as a false memory—could be formed. We first took virus-infected and fibre-implanted animals off Dox to open a time window for labelling cells activated by the exploration of a novel context (context A) with ChR2–mCherry. The animals were then immediately put back on Dox to prevent any further labelling. The next day, we fear-conditioned this group in a distinct context (context B) while optically reactivating the cells labelled in context A. In the following 2 days, we tested the animals’ fear memory in either the original context A or a novel context C.

If the light-reactivated cells labelled in context A can produce a functional CS during fear conditioning in context B, then the animals should express a false fear memory by freezing in context A, but not in context C.

Prior to conducting the behavioural experiments, we confirmed that distinct populations of cells in the DG represent contexts A and C (figure 4a–c), enabling the manipulation of context-specific memories at the level of defined neural populations. When DG cells activated by the exposure to context A were reactivated with light during fear conditioning in a distinct context B, the animals subsequently froze in context A at levels significantly higher than the background levels (figure 4f). Freezing in context C did not differ from background levels (figure 4f). This increased freezing in context A was not due to generalization because a control group

Figure 2. Basic experimental protocols and selective labelling of the DG cells by ChR2–EYFP. (a) The c-fos–tTA mouse was injected with AAVg–TRE-Chr2-EYFP and implanted with an optical fibre targeting the DG. (b) When off Dox, training induces the expression of c-fos–tTA, which binds to TRE and drives the expression of ChR2–EYFP, labelling a subpopulation of activated cells (yellow) in the DG. (c) Basic experimental scheme. Mice were habituated in context A with light stimulation while on Dox for 5 days, then taken off Dox for 2 days and fear conditioned (FC) in context B. Mice were put back on Dox and tested for 5 days in context A with light stimulation. (d) Representative image showing the expression of ChR2–EYFP in a mouse that was taken off Dox for 2 days and underwent fear conditioning training. An image of each rectangular area in (d) is magnified showing (e) DG, (f) CA1 and (g) CA3. The green signal from ChR2–EYFP in the DG spreads throughout granule cells, including dendrites (e), while the green signal confined to the nuclei in CA1 and CA3 is owing to shEGFP expression from the c-fos–shEGFP construct of the transgenic mouse (f,g). Blue is the nuclear marker DAPI.
expressing only mCherry that underwent the exact same training protocol did not show the same effect (figure 4f). These results indicate that artificial reactivation of DG cells initially activated by exposure to a particular context (context A) can serve as a functional CS during fear conditioning in a distinct context (context B) and results in the formation of a false memory [21].

4. No false-memory inception by CA1 engram manipulation

It is possible that not all c-Fos-expressing brain regions are sufficient to elicit the recall of a CS. Indeed, the hippocampus processes mnemonic information by shifting the combined activity of subsets of cells within defined subregions in response to discrete episodes [19,22]. Given that each subregion differentially contributes to an overall memory, we investigated whether a false memory could be formed by applying the same parameters and manipulations to CA1 as we did to the DG. Similar to the DG (figure 4a–e), the overlap was significantly lower across contexts (A and C) compared with a re-exposure to the same context (A and A) in CA1 (figure 4g–k). However, the degree of overlap for the two contexts was much greater in CA1 (30%) than in the DG (approx. 0.4%; figure 4e,k). Notably, exposure to a single context (A or C) consistently labelled approximately 50% of CA1 cells compared with approximately 6% of DG cells. When we labelled CA1 cells activated in context A and reactivated these cells with light during fear conditioning in context B, no increase in freezing was observed in the experimental group expressing ChR2–mCherry compared with the mCherry-only control group in either context (figure 4f). These data suggest that applying the same behavioural and stimulation parameters as we did in the DG does not result in the inception of a false memory in CA1.

5. Competition between false and genuine memory

According to classical learning theory, the simultaneous acquisition of two CSs can sometimes be a competitive phenomenon such that a memory for a single CS is acquired optimally when it is presented alone, whereas the presentation of two simultaneous CSs can lead to an overall decrement in behavioural output [23]. In our experiments, it is possible that the light-activated DG cells encoding context A interfered with the acquisition or expression of the genuine fear memory for context B. Indeed, upon re-exposure to context B, the experimental group froze significantly less than the group that did not receive light during fear conditioning or the group expressing mCherry alone (figure 5a). During light-on epochs in the context B test, freezing increased in the experimental group and...
decreased in the group that did not receive light during fear conditioning (figure 5a). We conducted similar experiments with mice in which the manipulation was targeted to the CA1 region and found no differences in the experimental or control groups during either light-off or light-on epochs of the context B test.

6. The false memory is a real memory

We could now probe the behavioural relevance of the DG cells that were artificially associated with an aversive event of high valence (e.g. US). If an artificial CS–US association was generated using our experimental parameters, then activation of the DG cells in a similar way as a genuine fear memory can be artificially recalled by light-reactivation of the corresponding output (i.e. freezing). To test whether a false fear memory can be artificially recalled by light-reactivation of the corresponding DG cells in a similar way as a genuine fear memory can be light-activated [16], we examined fear-memory recall in experimental and control groups of mice in a distinct context (context D) by administering light-off and light-on epochs (figure 5b). All groups exhibited background levels of freezing during light-off epochs. The experimental group, however, froze at significantly higher levels (approx. 25%) during light-on epochs. This light-induced freezing in context D was not observed in control animals that underwent the same behavioural schedule but did not receive light during fear conditioning in context B, in animals expressing mCherry alone, or in animals in which CA1 was manipulated instead (figure 5b).

To map the downstream brain areas involved during false-memory recall, in separate groups of animals we also performed histological analyses measuring c-Fos expression during three sessions: the false-memory recall test in context A, the context B natural fear-memory recall session and the context C control session. We predicted that the recall of a false memory engages neural substrates known to underlie genuine fear-memory recall, and therefore quantified the levels of c-Fos expression in the basolateral amygdala (BLA) and central amygdala (CeA) [24–29]. Indeed, false- and genuine-memory recall sessions elicited a significant increase in c-Fos-positive cells in the BLA and CeA compared with a control group exploring a neutral context (figure 5c–f).

These results indicated that similar circuit mechanisms underlie false memory as they do genuine-memory recall.

7. Discussion

Our data indicate that hippocampal DG cells activated previously during context exploration can subsequently serve as a functional CS in a fear-conditioning paradigm when artificially reactivated during the delivery of a US. The
Bonferroni posthoc test). and presence of light stimulation (**p = 0.001; #p = 0.034, two-way ANOVA followed by Bonferroni posthoc test). The percentage of c-Fos-positive cells was calculated for each group in BLA and CeA (**p = 0.007, two-way ANOVA followed by Bonferroni posthoc test). (c) Three groups of mice underwent the training shown in (a) and were sacrificed after testing in either context B (natural recall), A (false recall) or C (neutral context). The percentage of c-Fos-positive cells was calculated for each group in BLA and CeA (n = 6 subjects each; ***p < 0.001). Representative images for natural recall, false recall or neutral context are shown in (d), (e) and (f), respectively.

Figure 5. The false- and genuine-fear memories interact with each other and both recruit the amygdala. (a) Animals that underwent the behavioural protocol shown in figure 2g were re-exposed to context B and the freezing levels were examined both in the absence and presence of light stimulation (n = 11 for ChR2–mCherry group and n = 9 for ChR2–mCherry, no light group; *p = 0.027; ***p < 0.001; #p = 0.034, two-way ANOVA followed by Bonferroni posthoc test). (b) Animals that underwent the behavioural protocol shown in (a) were placed in a novel context D and the freezing levels were examined both in the absence and presence of light stimulation (n = 11 for ChR2–mCherry group and n = 9 for ChR2–mCherry, no light group; **p = 0.007, two-way ANOVA followed by Bonferroni posthoc test). (c) Three groups of mice underwent the training shown in (a) and were sacrificed after testing in either context B (natural recall), A (false recall) or C (neutral context). The percentage of c-Fos-positive cells was calculated for each group in BLA and CeA (n = 6 subjects each; ***p < 0.001). Representative images for natural recall, false recall or neutral context are shown in (d), (e) and (f), respectively.

consequence is the encoding of an artificial associative fear memory to the CS that was not naturally available at the time of the US delivery [30].

Memory is reconstructive in nature; the act of recalling a memory renders it labile and highly susceptible to modification [10–14]. In humans, memory distortions and illusions occur frequently. These modifications often result from the incorporation of misinformation into memory from external sources [10,11,13,14]. Interestingly, cognitive studies in humans have reported robust activity in the hippocampus during the recall of both false and genuine memories [15]. However, human studies using behavioural and fMRI techniques have not been able to delineate the hippocampal subregions and circuits that are responsible for generating false memories. To help to resolve these issues, our experiments provide an animal model in which false and genuine memories can be investigated at the memory engram level [31]. We propose that optical reactivation of cells that were naturally activated during the formation of a contextual memory induced the retrieval of that memory and, more importantly, the retrieved memory became associated with an event of high valence (i.e. a foot shock) to form a new but false memory that never had its component experiences naturally linked. Thus, the experimental group of animals showed increased freezing in a context in which they were never shocked (context A). Although our design for the formation and expression of a false memory was for a laboratory setting, and the retrieval of the contextual memory during conditioning occurred by artificial means (i.e. light), we suggest that the formation of at least some false memories may occur in natural settings by internally driven retrieval of previous experiences and their association with external stimuli of high valence.

A previous study applied a similar experimental protocol with pharmacosynthetic methods and failed to see increased freezing upon re-exposure to either context A or context B. Instead, they observed a synthetic memory that could only be retrieved by the combination of both contexts A and B [32]. A key difference in their system is that the c-Fos-expressing cells in the entire forebrain were labelled and reactivated over an extended period by a synthetic ligand. Moreover, the discrepancy observed here suggests two methodological caveats: the spatial and millisecond precision of region-specific optogenetic manipulations, when compared to forebrain-wide pharmacogenetic perturbations that last several minutes, perhaps more reliably recapitulates
the endogenous neural activity required for behavioural expression of a memory, and perhaps not all c-Fos-expressing brain regions are sufficient to elicit the recall of a CS. We propose that activating neurons in much wider spatial and temporal domains may favour the formation of a synthetic memory, which may not be easily retrievable by the cues associated with each individual memory. By contrast, activating neurons in a more spatially (only small populations of DG cells) and temporally restricted manner (only a few minutes during light stimulation) may favour the formation of two distinct (false and genuine) memories as observed in our case. In line with this hypothesis, when we manipulated CA1 cells by the same procedures as the ones used for DG cells, we could not incept a false memory (i.e. freezing in context A). In CA1, the overlap of the cell populations activated by consecutive exposures to a pair of contexts is much greater than that in the DG. We hypothesize that our negative CA1 behavioural data could be a result of contextual engrams relying less on a population code and increasingly on a temporal code as they travel through the trisynaptic circuit [8,9,22].

Together, our findings provide a foundation for an experimental bridge between the traditions of rodent behavioural neuroscience and human cognitive neuroscience in the study of memory by illuminating the underlying circuits supporting internally generated representations and their contribution to the formation of false memories.

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**References**