Ca\textsuperscript{2+} toxicity and mitochondrial damage in acute pancreatitis: translational overview

József Male\textethetics\textsuperscript{1,2} and Péter Hegyi\textsuperscript{1,2,3}

\textsuperscript{1}First Department of Medicine, and \textsuperscript{2}MTA-SZTE Momentum Translational Gastroenterology Research Group, University of Szeged, Szeged, Hungary
\textsuperscript{3}Institute for Translational Medicine, University of Pécs, Pécs, Hungary

Acute pancreatitis (AP) is a leading cause of hospitalization among non-malignant gastrointestinal disorders. The mortality of severe AP can reach 30–50%, which is most probably owing to the lack of specific treatment. Therefore, AP is a major healthcare problem, which urges researchers to identify novel drug targets. Studies from the last decades highlighted that the toxic cellular Ca\textsuperscript{2+} overload and mitochondrial damage are key pathogenic steps in the disease development affecting both acinar and ductal cell functions. Moreover, recent observations showed that modifying the cellular Ca\textsuperscript{2+} signalling might be beneficial in AP. The inhibition of Ca\textsuperscript{2+} release from the endoplasmic reticulum or the activity of plasma membrane Ca\textsuperscript{2+} influx channels decreased the severity of AP in experimental models. Similarly, inhibition of mitochondrial permeability transition pore (MPTP) opening also seems to improve the outcome of AP in in vitro animal models. At the moment MPTP blockers are under detailed clinical investigation to test whether interventions in MPTP openings and/or Ca\textsuperscript{2+} homeostasis of the cells can be specific targets in prevention or treatment of cell damage in AP.

This article is part of the themed issue ‘Evolution brings Ca\textsuperscript{2+} and ATP together to control life and death’.

1. Ca\textsuperscript{2+} is controlling secretory events in pancreatic acinar and ductal cells

Intracellular Ca\textsuperscript{2+} signalling plays central role in the regulation of the secretory processes of the exocrine pancreas. It is a well-known fact that in the exocrine pancreas acinar cells secrete digestive enzymes and pancreatic ductal epithelial cells secrete HCO\textsubscript{3} rich alkaline fluid that washes the digestive enzymes out from the pancreas. The prompt coordination of the secretory events of the two cell types is essential and Ca\textsuperscript{2+} has a central role in both pancreatic physiology and pathophysiology. Recent studies suggest that these two cell types cannot be handled separately since they are more likely integrated into a functional unit [1]. This is further amplified by the neurohormonal regulation of exocrine pancreatic secretion. It has been demonstrated that acetylcholine (the main stimulatory neurotransmitter of the pancreas) is released from the parasympathic nerve endings, releasing digestive enzymes from the acinar cells [2], whereas at the same time enhances the pancreatic ductal fluid and HCO\textsubscript{3} secretion via M\textsubscript{3} metabotropic cholinergic receptor (M3R) mediated Ca\textsuperscript{2+} release [3]. In addition, the circulating hormone cholecystokinin (CCK) regulates pancreatic secretion via oscillatory Ca\textsuperscript{2+} signals [4]. In pancreatic ductal epithelial cells (PDECs), the role of CCK stimulation differs between species, in humans it has negligible direct effects, but remarkably potentiates the stimulatory effect of secretin on the HCO\textsubscript{3} secretion [5]. The proper control of secretion is further potentiated by the strong synergy between Ca\textsuperscript{2+} and cAMP signalling [6]. The physiological roles of...
Ca\(^{2+}\) signalling in epithelial secretion have been outlined in more detail in excellent reviews [7–10].

2. The price of versatility: Ca\(^{2+}\) toxicity in acute pancreatitis

Although it is well established that physiological Ca\(^{2+}\) signalling controls the normal pancreatic functions on multiple levels, it is also well documented that uncontrolled cellular Ca\(^{2+}\) overload leads to cellular damage and pathogenesis of acute pancreatitis (AP; figure 1). In this chapter, we will summarize the effects of the common stress factors that cause AP.

(a) Bile acids

Biliary pancreatitis is one of the most common forms of AP, although the exact pathogenesis is not known in detail. One possible explanation is the ‘common channel’ theory, which suggests that an impacted gallstone creates communication behind the stone connecting the common bile duct to the pancreatic duct. This would theoretically allow bile acids (BAs) to reach the pancreatic ductal lumen or even the acinar cells [11]. However, this hypothesis was questioned by several studies suggesting that instead of the reflux mechanism, increased luminal pressure would cause the pancreatic damage [12–15]. Whether or not BAs reach the pancreatic tissue directly from the luminal side, several observations suggest that BA reaching the ductal cells from either basolateral or luminal sides can trigger multiple cellular responses in acinar and ductal cells that might contribute to the development of AP.

Earlier, our group showed that the hydrophobic, non-conjugated BA, chenodeoxycholate (CDCA) dose-dependently affects \(\text{HCO}_3^-\) secretion of pancreatic ductal epithelia [16]. We found that lower concentration of CDCA (100 \(\mu\)M) stimulated and high concentration (1 mM) severely inhibited the ion transport activities including the ductal \(\text{HCO}_3^-\) secretion. The explanation for this dual effect might be the type of Ca\(^{2+}\) signals triggered by CDCA. Luminal administration of 100 \(\mu\)M CDCA evoked short oscillatory Ca\(^{2+}\) signals, which were fully abolished by IP\(_3\) receptor inhibition. On the other hand, challenging the pancreatic ductal cells with 1 mM CDCA caused a sustained Ca\(^{2+}\) elevation [16] and severe damage of the mitochondrial morphology and function [17]. Interestingly, in our hands N,N\'-[1,2-ethanediybis(oxy-2,1-phenylene)]bis[N-[2-[(acetyloxy)methoxy]-2-oxoethy]l]-bis[acetyloxy]methyl]ester (BAPTA-AM) failed to prevent the mitochondrial damage and therefore the inhibitory effect of CDCA on the \(\text{HCO}_3^-\) secretion [16], which might be explained by the existence of a Ca\(^{2+}\)-independent direct mitochondrial toxicity of bile acids [18]. Similarly to ductal cells, pancreatic acinar cells respond with intracellular Ca\(^{2+}\) elevation to BA challenge [19] due to IP\(_3\)R and ryanodine receptor activation. It is also well documented that tauroliothiocholic acid 3-sulfate diminishes cellular ATP production [20] and dissipate the mitochondrial membrane potential (\(\Delta \Psi_m\)), which was not affected by BAPTA-AM treatment [21]. Although BA directly affects the acinar cells, the observations of Perides et al. actually suggest that biliary pancreatitis is a receptor mediated disease [22]. They showed that the G-protein-coupled cell surface bile acid receptor (Gpbar1, or TGR5) is expressed at the apical membrane of pancreatic acinar cells and its activation is associated with pathological Ca\(^{2+}\) signals, intracellular activation of digestive enzymes and cell injury, i.e. the hallmarks of AP. Whereas the genetic deletion of Gpbar1 markedly reduced the severity of tauroliothiocholic acid 3-sulfate (TLCS)-induced, but not caerulein-induced AP. Very recently, Katona et al. provided solid evidence that specific BA might be used as treatment option against biliary pancreatitis [23]. They showed that pre-treatment of pancreatic ducts with ursodeoxycholate (UDCA) remarkably ameliorated the toxic effects of UDCA. Chenodeoxycholate-induced intracellular
ATP depletion, mitochondrial injury, and as a consequence, cell death were completely prevented by UDCA, whereas the activity of the epithelial cell–base transporters was preserved in *in vitro* experiments. In addition, *in vivo* experiments showed that oral administration of UDCA significantly reduced the severity of CDCA-induced AP. Interestingly, UDCA had no effect on the sustained \( \text{Ca}^{2+} \) elevation triggered by CDCA, raising the possibility of a direct mitochondrial protective effect, which is yet to be determined. These observations nicely supplement the previous results of Seyhun *et al.*, who showed that the endoplasmic reticulum (ER) chaperone tauroursodeoxycholic acid inhibits the unfolded protein response (UPR) *in vitro* [24] and *in vivo* [25]. This effect reduced the activation of UPR components and reduced intracellular trypsin activation, oedema formation and cell damage in pancreatic acinar cells.

**(b) Ethanol and non-oxidative ethanol metabolites**

The second most frequent form of pancreatitis is alcohol-induced AP [26]. Whereas genetic factors seem to be involved in the disease development [27], several studies investigated the direct effects of ethanol and different ethanol metabolites on the exocrine pancreas. Ethanol and its oxidative metabolite acetaldehyde have moderate effects on the \( \text{Ca}^{2+} \) in pancreatic acinar cells even in extremely high concentrations [28]. Whereas the non-oxidative ethanol metabolites (fatty acid ethyl esters, FAEE) induced sustained \( \text{Ca}^{2+} \) elevation and a drop of cellular ATP leading to necrosis [28–30]. Importantly, the breakdown of FAEE to fatty acids (FA) by intracellular hydrolases significantly contribute to the toxic effects of non-oxidative ethanol metabolites [30]. This fact has been further emphasized in a recent elegant study by Huang *et al.* [31]. They showed that the inhibition of oxidative ethanol metabolism significantly enhance, whereas inhibition of non-oxidative ethanol metabolism augment pancreatic damage in an *in vivo* model of ethanol-fatty acid induced AP. On the other hand pancreatic ductal cells respond to low to high concentrations of alcohol, likewise to BA. Yamamoto *et al.* showed that 1 mM ethanol induces \( \text{Ca}^{2+} \) elevation and augments fluid secretion, whereas high concentration moderately inhibits the stimulated fluid secretion in secretin-stimulated guinea pig pancreatic ducts [32]. Our group recently investigated the effects of ethanol and ethanol metabolites in more detail [33]. We showed that alcohol and fatty acids inhibit fluid and \( \text{HCO}_3^- \) secretion, as well as cystic fibrosis transmembrane conductance regulator (CFTR) activity, in pancreatic ductal cells. Interestingly, in the case of FAEE only the inhibition of the CFTR channel was observed in high concentrations [34]; however, the inhibition of \( \text{HCO}_3^- \) secretion was not observed [33]. The remarkable inhibitory effects of alcohol and fatty acids were mediated by sustained increase of intracellular \( \text{Ca}^{2+} \), inhibited adenosine 3',5'-cyclic monophosphate and ATP production and depolarization of \( \Delta V_{m} \). We also showed that ethanol reduced expression of CFTR via multiple pathways, which in turn augmented the severity of experimental alcohol-induced AP in mice.

**(c) Other stress factors**

As demonstrated above, the two most common pathogenic factors of AP—BA and ethanol—damage the exocrine pancreas via \( \text{Ca}^{2+} \) toxicity and mitochondrial injury. Notably, these cellular changes seem to be the key of AP pathogenesis since a considerable number of studies showed that other stress factors provoke the same alterations in \( \text{Ca}^{2+} \) signalling and energy metabolism. Intrapancreatic trypsinogen activation is a hallmark of AP pathogenesis and we showed earlier that trypsin acting via PAR2 on the luminal membrane induces intracellular \( \text{Ca}^{2+} \) elevation and inhibits the luminal acid/base transporters in PDEC [35]. Moreover, the inhibitory effect was abolished by BAPTA-AM preincubation, similarly to the inhibitory effects of ethanol and fatty acids. Very recently, Jin *et al.* investigated the pathomechanism of an iatrogen form of AP, the post-ERCP pancreatitis [36]. Using sophisticated *in vitro* and *in vivo* models, they showed that exposure of pancreatic acinar cells to iohexol (a radiocontrast agent) triggered sustained intracellular \( \text{Ca}^{2+} \) elevation. The downstream activation of NF-κB and NFAT is completely abolished by the suppression of the \( \text{Ca}^{2+} \) signals. Moreover, they proved that the downstream effects of \( \text{Ca}^{2+} \) were mediated by calcineurin since genetic, or pharmacological inhibition of calcineurin prevented the radiocontrast-induced damage. This interesting study further underlines the central role of pathophysiological \( \text{Ca}^{2+} \) signalling in the pathogenesis of AP regardless of the etiological factor.

### 3. Sources of \( \text{Ca}^{2+} \) in pancreatic acinar and ductal cells

**(a) \( \text{Ca}^{2+} \) release from the endoplasmic reticulum**

Agnost binding (Ach, ATP) to G-protein-coupled receptors activate phospholipase C β (PLCβ) in pancreatic acinar and ductal cells. The activated PLCβ releases inositol trisphosphate (IP3) by hydrolysing phosphatidylinositol 4,5-bisphosphate (PIP2) [37]. Under physiological conditions, the intracellular \( \text{Ca}^{2+} \) signals have a strict spatio-temporal localization [38,39], mostly limiting \( \text{Ca}^{2+} \) signals to the apical pole of the cells. As in other non-excitable cell types, this is ensured by two ATP-dependent pumps that clear the cytosol from the free \( \text{Ca}^{2+} \). The sarco/endoplasmic reticulum \( \text{Ca}^{2+} \)-ATPase (SERCA) pumps and the plasma membrane \( \text{Ca}^{2+} \)-ATPase (PMCA) pumps move \( \text{Ca}^{2+} \) from the cytosol to the ER and the extracellular space, respectively. This activity restores basal intracellular \( \text{Ca}^{2+} \) levels and refills the ER \( \text{Ca}^{2+} \) stores. In PDEC, the \( \text{Ca}^{2+} \) signalling is not characterized in such detail; however, the overall polarity of the ductal cells including the ion channels and transporters, IP3 receptors and mitochondria [17], suggest a very similarly regulated \( \text{Ca}^{2+} \) signalling, like in acinar cells. Further studies are required for the clarification of these questions.

**(b) Extracellular \( \text{Ca}^{2+} \) influx**

The complex role of extracellular \( \text{Ca}^{2+} \) influx to orchestrate non-excitable cell functions has been established several decades ago [40]; however, the molecular components participating in the process remained unknown until 2005. Hoth *et al.* found that agonist-mediated depletion of the intracellular \( \text{Ca}^{2+} \) stores induced a \( \text{Ca}^{2+} \) selective sustained inwardly rectifying current, which was termed \( i_{\text{TRAC}} \) (calcium release-activated calcium current) [41]. The real revolution of the field began by the discovery of the ER \( \text{Ca}^{2+} \) sensor stromal interaction molecule 1 (Stim1) [42] and the plasma membrane \( \text{Ca}^{2+} \) channel Orai1 [43,44].Briefly, the process of store operated \( \text{Ca}^{2+} \) entry (SOCE) consist of the following elements. In resting conditions the ER \( \text{Ca}^{2+} \) stores are refilled and Stim1 distributes in the ER membrane. However during physiological stimulation the ER \( \text{Ca}^{2+} \) stores are quickly depleted, which induces the dissociation of the bound \( \text{Ca}^{2+} \)
from the EF hand of Stim1. This is followed by a conformational change and translocation of Stim1 to defined ER-PM junctions, termed as puncta formation [45]. This translocation is required for the activation of the plasma membrane Ca\(^{2+}\) influx channel Orai1, where the Stim Orai1-activating region (SOAR) and poly-basic domains of Stim1 interact with different binding sites of Orai1 that results in clustering and activation of the channel [46]. In addition to Orai1, other possible Ca\(^{2+}\) entry channels that seem to play a role in Stim1-mediated SOCE are the TRPC channels [47,48]. These channels function as Ca\(^{2+}\)-permeable non-selective cation channels mediating receptor evoked Ca\(^{2+}\) influx in many cells [49]. SOCE have been investigated mostly in acinar cells of various exocrine glands, as models of polarized epithelial cells [50–52]. Interestingly, the role of SOCE in the physiological functions of PDEC, especially in HCO\(_3^-\) secretion remained elusive. Kim et al. found that intracellular Ca\(^{2+}\) elevation, caused by the activation of SOCE might play a role in exocytosis in pancreatic ductal cells isolated from dog main pancreatic duct [53,54]; however, they did not investigate HCO\(_3^-\) secretion of PDEC, which therefore needs further investigation.

4. Mitochondrial Ca\(^{2+}\) handling and Ca\(^{2+}\) overload of mitochondria

During physiological Ca\(^{2+}\) signalling, mitochondria takes up Ca\(^{2+}\), which has been shown to directly increase energy output by enhancing the activity of tricarboxylic acid cycle dehydrogenases and the ATP synthase [55]. The pioneer work of Rizzuto et al. highlighted that the cytosolic Ca\(^{2+}\) signals propagate to the mitochondria [56] and a couple of years later Csurdas et al. found that ER membrane and the outer mitochondrial membrane form a quasi-synaptic connection [57] that is the structural bases of the Ca\(^{2+}\) hotspots [58]. Despite the functional characterization of the mitochondrial Ca\(^{2+}\) signalling the molecular background of the process was not known. In 2011, two groups independently identified the mitochondrial Ca\(^{2+}\) uniporter (MCU), an inner mitochondrial membrane protein that is responsible for the mitochondrial Ca\(^{2+}\) uptake [59,60]. The Ca\(^{2+}\) efflux from the mitochondria is mediated by the mitochondrial Na\(^+\)/Ca\(^{2+}\) exchanger (NCLX) [61], thus the mitochondrial Ca\(^{2+}\) level is tightly regulated under physiological conditions. However, pathophysiological signals can lead mitochondrial injury, which can activate both apoptosis and necrosis. The classical mitochondrial apoptotic pathway involves the outer membrane permeabilization by Bax and Bak (two members of the pro-death Bcl-2 family) that will allow apoptotic factors like cytochrome c, Smac/DIABLO and apoptosis inducing factor to be released from the intermembrane space into the cytosol, leading to cell death by apoptosis [62]. On the other hand, Ca\(^{2+}\) overload or increased reactive oxygen species (ROS) production can cause the opening of mitochondrial permeability transition pore (MPTP) that results in the loss of mitochondrial inner membrane potential, uncoupling of the respiratory chain with a consequent drop of mitochondrial ATP synthesis, and increased permeability of the inner mitochondrial membrane that eventually leads to mitochondrial swelling, rupture and necrotic cell death [63,64]. Notably, recent studies lead to the reconsideration of the role of MPTP in cellular physiology, since it has been proved to be important in several physiological processes such as energy metabolism [65], mitochondrial Ca\(^{2+}\) efflux [66] and ROS signalling [67] as well. The molecular identity of MPTP is still a matter of investigation [68]. The historical model of MPTP included the voltage-dependent anion channel (VDAC) in the outer mitochondrial membrane, the adenine nucleotide translocator (ANT) in the inner mitochondrial membrane, and CypD as its regulator in the matrix of the mitochondria [69]. However recent intensive efforts revealed new molecules that might contribute to the MPTP formation (reviewed in detail [68,70]). A growing number of evidence suggest that VDAC is not very likely to contribute to the MPTP formation. On the other hand, studies on ANT suggest that it is not required for MPTP formation, but it regulates MPTP activity [71]. CypD is an important regulator of MPTP as supported by genetically modified mice [72] and pharmacologic inhibition of CypD by cyclosporine A [73]. On the other hand, several studies suggested that the activity of F1F0 ATP synthase or the proapoptotic Bax/Bak proteins [74] are required for proper MPTP function [75], whereas other proteins, such as mitochondrial phosphate carrier, might impact the pore opening indirectly [76]. At the moment the role of MPTP in the pathogenesis of AP is supported by limited, but still solid evidence. Mukherjee et al. demonstrated that both genetic and pharmacologic inhibition of MPTP opening (using the Cyclophilin D-deficient Pppf gene knockout mice, or in vivo treatment with cyclosporine A derivate) significantly ameliorated pancreatic damage in different experimental AP models in mice [77]. Importantly MPTP blockade protected the pancreatic acinar cells from necrosis whereas apoptosis was not affected, which is in strong agreement of earlier studies [72].

5. Novel therapeutic targets in acute pancreatitis

In pancreatic acinar cells, IP\(_{\gamma}\)-mediated Ca\(^{2+}\) release from the ER is an essential component of the physiological response to agonist stimulation, but it could also contribute to the pathological Ca\(^{2+}\) overload of the cells evoked by toxic factors that induce AP (cerulin hyperstimulation, bile acids, or ethanol and ethanol metabolites) [39]. Caffeine is a known inhibitor of IP\(_{\gamma}\)Rs due to multiple actions that include the inhibition of phospholipase C-mediated production of IP\(_3\) [78], antagonism of IP\(_{\gamma}\)Rs [79] and direct binding to IP\(_{\gamma}\)s that reduce the channels open-state probability [80]. Interestingly, coffee consumption moderately reduces the risk of alcohol-associated pancreatitis suggesting that the inhibitory effect of caffeine on IP\(_{\gamma}\)-mediated Ca\(^{2+}\) signalling may be protective in AP [81]. Based on these considerations Huang et al. recently studied the effects of caffeine and its xanthine metabolites on pancreatic acinar IP\(_{\gamma}\)-mediated Ca\(^{2+}\) signalling and experimental AP [82]. They found that caffeine and dimethylxanthines (but not monomethylxanthines) blocks IP\(_{\gamma}\)-mediated Ca\(^{2+}\) oscillations in response to uncaged IP\(_3\) or toxins, prevented mitochondrial depolarization and necrotic cell death in vitro and significantly impaired the severity of experimental AP in three different models. These observations suggest that caffeine, or its metabolites might be suitable starting points to develop therapy for AP (figure 2).

As discussed above, store operated Ca\(^{2+}\) entry could be a key component in the development of cellular Ca\(^{2+}\) overload. Earlier Kim et al. showed that genetic [83] or pharmacological inhibition (using the TRPC3-specific inhibitor pyrazole 3) [84] of TRPC3 significantly reduce the sustained Ca\(^{2+}\) elevation in pancreatic acinar cells evoked by cell stressors (bile acid or fatty acid ethyl ester). In addition, it prevented the pathological
inhibition of digestive enzyme secretion and markedly reduced intracellular trypsin activation and excessive actin depolymerization in vitro and the severity of pancreatitis in vivo. Recently, Gerasimenko et al. demonstrated the pharmacological inhibition of another Ca^{2+} entry channel Orai1 by a specific inhibitor called GSK-7975A which prevents acinar cell necrosis in vitro [85]. This important observation was supported by Wen et al., who tested the effects of two specific Orai1 inhibitors (GSK-7975A and CM_128) in isolated human and rodent pancreatic acinar cells and in different experimental AP models [86]. They showed that both Orai1 inhibitors prevented the sustained Ca^{2+} elevation in vitro and significantly impaired signs of pancreatic injury including pancreatic oedema, inflammation and necrosis in all tested experimental models.

Mitochondrial permeability transition is a key feature of cellular damage in many cell types and diseases (see above); therefore, MPTP blockers are under detailed clinical investigation in different studies. In a recent clinical study, the efficacy and safety of TRO40303 (an MPTP inhibitor) have been evaluated for the reduction of reperfusion injury in patients undergoing revascularization for ST-elevation myocardial infarction (MITOCARE study) [87]. This study did not show any effect of TRO40303 in limiting reperfusion injury of the ischaemic myocardium. In another recently completed CIRCUS trial, the effects of i.v. administered cyclosporine have been evaluated on the clinical outcome of patients with anterior STEMI [88]. Similarly to the MITOCARE study, CIRCUS trial did not report any improvement in the cyclosporine-treated patients. The reasons for the failure of the studies might be explained by pharmacological limitations of the administered compounds [89] that include low tissue penetration due to the lack of collateral blood flow and high metabolism of the compound in the blood. In addition, MPTP blockers have been suggested to be beneficial in hepatitis C therapy, since they inhibited hepatitis C virus (HCV) replication by preventing a cyclophilin-A induced cis–trans isomerization in domain II of NS5A [90]. However, it was not investigated in clinical trials further. Very recently, Mukherjee et al. tested the effect of MPTP inhibition on the severity of AP in rodent experimental AP models [97]. They have shown that the inhibition of MPTP with pharmacological compounds (two cyclosporin A derivate: DEB025 or TRO40303), or genetic deletion of the Ppif gene (that encodes cyclophilin D, a component of MPTP) significantly decreases the severity of AP in different independent models. These observations suggest that the MPTP inhibition might be potentially beneficial in the AP therapy. Other indirect evidence for this hypothesis has been provided by Judak et al., who showed that the supplementation of cellular ATP in vitro diminished the inhibitory effect of ethanol metabolites on the ion transport activities in isolated guinea pig pancreatic ductal cells [34]. These results suggest that the restoration of the cellular energy level can be beneficial in AP, which can prevent the cellular dysfunction and cell damage.

6. Closing remarks

Although there are several promising results and potential drug targets that play a role in the pathogenesis of AP, it remains a great challenge for researchers and clinicians. A number of unanswered questions are waiting for answers. Moreover, it will take several years to test the experimental results on clinical patients as well. To be able take up these challenges, clinicians and researchers should work closely together in the future.

Competing interests: We declare we have no competing interests.

Funding. Our research was supported by the Hungarian Scientific Research Fund (PD119974 (J.M.), K116634 (P.H.)) and the Momentum grant of the Hungarian Academy of Sciences (LP2014-10/2014 (P.H.)).

References


6. Ahuja M, Jha A, Maleth J, Park S, Mualem S. 2014 cAMP and Ca(2+)


89. Coelmont L et al. 2010 DEB025 (Alisporivir) inhibits hepatitis C virus replication by preventing a cyclophilin A induced cis-trans isomerisation in domain II of NS5A. PLoS ONE \textbf{5}, e13687. (doi:10.1371/journal.pone.0013687)